

The role of labile Zn²⁺ and Zn²⁺-transporters in the pathophysiology of mitochondria dysfunction in cardiomyocytes

Belma Turan^{1,2} • Erkan Tuncay²

Received: 27 August 2020 / Accepted: 23 October 2020 / Published online: 22 November 2020 © Springer Science+Business Media, LLC, part of Springer Nature 2020

Abstract

An important energy supplier of cardiomyocytes is mitochondria, similar to other mammalian cells. Studies have demonstrated that any defect in the normal processes controlled by mitochondria can lead to abnormal ROS production, thereby high oxidative stress as well as lack of ATP. Taken into consideration, the relationship between mitochondrial dysfunction and overproduction of ROS as well as the relation between increased ROS and high-level release of intracellular labile Zn²⁺, those bring into consideration the importance of the events related with those stimuli in cardiomyocytes responsible from cellular Zn²⁺-homeostasis and responsible Zn²⁺-transporters associated with the Zn²⁺-homeostasis and Zn²⁺-signaling. Zn^{2+} -signaling, controlled by cellular Zn^{2+} -homeostatic mechanisms, is regulated with intracellular labile Zn^{2+} levels, which are controlled, especially, with the two Zn²⁺-transporter families; ZIPs and ZnTs. Our experimental studies in mammalian cardiomyocytes and human heart tissue showed that Zn²⁺-transporters localizes to mitochondria besides sarco(endo) plasmic reticulum and Golgi under physiological condition. The protein levels as well as functions of those transporters can re-distribute under pathological conditions, therefore, they can interplay among organelles in cardiomyocytes to adjust a proper intracellular labile Zn²⁺ level. In the present review, we aimed to summarize the already known Zn²⁺-transporters localize to mitochondria and function to stabilize not only the cellular Zn²⁺ level but also cellular oxidative stress status. In conclusion, one can propose that a detailed understanding of cellular Zn²⁺-homeostasis and Zn²⁺-signaling through mitochondria may emphasize the importance of new mitochondria-targeting agents for prevention and/or therapy of cardiovascular dysfunction in humans.

Keywords Zinc · Heart · Hyperglycemia · Hyperinsulinemia · Aging · Mitochondria · Zinc-transporters

Introduction

Mitochondria, similar to most mammalian cells, occupy the large part of a cardiomyocyte and play vital roles in alive cells. Under physiological conditions, mitochondria mainly function to provide the required energy to the beating heart via producing ATP through oxidative phosphorylation [1–7]. Therefore, those abundant mitochondria maintain the energy need of cells, as a perfect ATP source, to support contraction, metabolism, and ion homeostasis in cardiomyocytes.

☑ Belma Turan belma.turan@medicine.ankara.edu.tr; belma.turan@lokmanhekim.edu.tr Since cell metabolic activity besides energy is derived from mitochondria under physiological conditions, therefore, mitochondrial dysfunction is considered to be a therapeutic target for pathological conditions including cardiac dysfunction [8]. Any abnormalities in mitochondrial fission—fusion dynamics (i.e. altered expression of mitochondrial proteins) and bioenergetics can lead to cardiovascular diseases [9, 10]. In other words, mitochondrial dysfunction, including structural and metabolic alterations, contributes to heart diseases besides others.

Studies pointed out that oxidative stress is the main molecular mediators of heart diseases in patients and experimental animals while these mediators regulate both the degradation and remodeling processes in the heart [7, 11]. In that regard, it has been shown that not only reactive oxygen species (ROS) but also reactive nitrogen species (RNS) play important in the development of cellular abnormalities such as defective Ca²⁺-handling (causing cardiac arrhythmia) as



Department of Biophysics, Faculty of Medicine, Lokman Hekim University, Ankara, Turkey

Department of Biophysics, Faculty of Medicine, Ankara University, Ankara, Turkey

well as inducing hypertrophic signaling, apoptosis, and necrosis [12–15]. Often, these alterations are caused by genetic mutations in mitochondrial DNA [16]. In line with that statement, now, it is also well known that mitochondrial dysfunction and associated ROS over-generation lead mainly to extensive oxidative stress and less ATP production, which in turn causes the activation of mitochondrial-driven cell death via the opening of mPTP [8, 17, 18].

We, previously, have shown that Zn²⁺ is releasing into the cytosol during the cardiac excitation-contraction cycle in a manner of both Ca²⁺ and redox-dependent and can trigger ROS production via inducing changes in metal-binding properties of metallothioneins [19, 20]. Furthermore, over ROS production can induce a high level of intracellular Zn²⁺ releases under pathological stimuli such as hyperglycemia and/or exposure directly to oxidants [21–25]. Indeed, we demonstrated that disturbances in cellular Zn²⁺ levels in cardiomyocytes could contribute and/or exacerbate heart dysfunction observed under chronic hyperglycemic conditions [18, 26–28].

It has been also shown that a significant increase in intracellular free Zn²⁺ could induce marked increases in mitochondrial matrix/cristae area and matrix volume together with increased lysosome numbers in mammalian cardiomyocytes. Also, there were notable clustering and vacuolated mitochondrion markedly disrupted and damaged myofibrils and electron-dense small granules with some implications of fission-fusion defects in the mitochondria in those cells [18, 26]. In terms of functional changes in those Zn²⁺ exposed cardiomyocytes, there was marked depolarization in mitochondrial membrane potential as well as a high level of ROS production [28, 29]. Those findings are highlighting the close association between cellular free Zn²⁺ level, oxidative stress, and mitochondrial function in cardiomyocytes under not only pathological stimuli but also for their physiological function.

Therefore, a better understanding of this cellular crosstalk might help to develop new ways to prevent and/or treat heart diseases. Under the light of this hypothesis, here, we aimed to document and discuss the current data in this subject.

Labile Zn²⁺ plays an important role in the regulation of cardiac cell function

Both experimental and clinical studies demonstrate that impairment of Zn^{2+} -homeostasis leads to alterations in the body which leads to induce a variety of health problems [30–32]. Among them, zinc-deficiency can affect human health, including cardiovascular function among others [33–35]. However, there are some controversies related to the labile Zn^{2+} role in mammalian cells, particularly in

cardiomyocytes, such as its opposing effects. The recent and early studies indicate that Zn^{2+} is a co-factor for several enzymes in the antioxidant defense system, thereby, protects cells against oxidative damage [31, 36–41]. Also, Zn^{2+} acts in the stabilization of membranes inhibit the enzyme nicotinamide adenine dinucleotide phosphate oxidase (NADPH-Oxidase), a pro-oxidant enzyme, and induces metallothionein synthesis [42]. However, studies also emphasized that elevated intracellular labile Zn^{2+} is toxic for cardiomyocytes similar to those of other cells, through essentially its action on the modulation of protein gene expression and mitochondrial and SER functions [26, 28, 29, 43–45].

Correspondingly, it is reported that an optimal ratio of labile Zn²⁺ level to labile Ca²⁺ level in cytosol and mitochondria can be preserved to combat oxidative stress by the protection of cardiomyocyte-injury by different stimuli including high Zn²⁺ through a well-controlled mitochondrial function [46–49]. Of note, it has been previously shown that the total intracellular labile Zn²⁺ level in ventricular cardiomyocytes is less than 1-nM in both rat and rabbit ventricular cardiomyocytes under physiological conditions [45, 50, 51]. Under pathological conditions, including hyperglycemia, hyperinsulinemia, and aging as well as acute oxidant exposures, its level can increase either over twofold or 30-fold [19, 20, 25, 29, 45, 48, 50]. Together, it should be emphasized that there are important cellular toxicity of high intracellular labile Zn²⁺ in cardiomyocytes and this type of toxicity can in turn lead to the Ca²⁺ dyshomeostasis, impairment in excitation-contraction coupling as well as mitochondrial dysfunction. These alterations will result from important elevation in the production of ROS and/or RNS, apoptosis, and cell death in cells including cardiomyocytes [19, 26, 28, 39, 45, 52–56]. Although the exact molecular mechanisms of high intracellular labile Zn²⁺ toxicity in cells, its interactions with cysteinyl thiols of proteins thereby its participation in the redox reactions seems to be at most its molecular effect in ventricular cardiomyocytes [21, 26]. Furthermore, in our previous studies performed in heart preparations, we have shown that all these toxic changes and damages via high intracellular labile Zn²⁺ in tissue and cell levels were at most associated with increases in not only ROS but also RNS levels. Correspondingly, the light and electron microscopy examinations of cardiomyocytes incubated exposed to high Zn²⁺ demonstrated clear hypertrophy in cardiomyocytes, and increased numbers of lysosomes and lipid droplets in the interstitial area, besides markedly disrupted and damaged myofibrils [18, 26]. Therefore, it seems that intracellular high Zn²⁺ toxicity is closely associated with increased oxidative stress, while increased oxidative stress can induce further increase in intracellular labile Zn²⁺ through Zn²⁺ release from subcellular stores [28, 45, 57]. Altogether, one can propose that increased intracellular Zn²⁺ is leading to



the induction of deleterious changes to stimulate different cardiac dysfunction [25, 28, 57, 58].

Two faces of zinc in biological systems: Zinc and oxidative stress

Zinc is not only a co-factor for many enzymes involved in the physiological role of the antioxidant defense system but also protects cells against oxidative damage through stabilizing the homeostasis of several intracellular pathways. Among its activities, it plays an important role in restoring impaired energetic metabolism via the stabilization of membranes, ionic homeostasis as well as it mediates the phosphorylation and oxidation of several proteins, kinases, and enzymes [25, 59, 60]. Studies also have shown that it plays an important role in the conversion of two superoxide radicals to hydrogen peroxide and molecular oxygen, reducing the toxicity of ROS [61]. However, we and others demonstrated its toxic effect that an increase in intracellular labile Zn²⁺ level can elevate in cardiomyocytes by ROS/RNS through in a process dependent on Zn²⁺ release from intracellular stores [31, 45, 53, 62]. Correspondingly, through the contribution of elevated ROS/RNS to the damage and dysfunction in cardiomyocytes, one can interpret why there is a close relationship between increased intracellular labile Zn²⁺ level and deleterious changes in several signaling pathways in the heart [18, 21, 25, 26, 28, 45, 53, 62].

Similar to the intracellular Ca²⁺-homeostasis, the intracellular Zn²⁺-homeostasis is dynamically maintained by a variety of proteins, kinases, and enzymes as well as sharing the same intracellular stores which are distributed in distinct cellular compartments of cardiomyocytes [9, 19, 47, 57, 63]. Those actors responsible for the homeostasis, are very sensitive to increased oxidative stress in cell levels.

Although Zn²⁺ itself is not a direct redox-active element, it plays an important and complex interplay in many cells including cardiomyocytes [45]. It has been shown modulation of intracellular labile Zn²⁺ level in cells by the redox state (i.e. increased ROS) [64]. Together with that property, it increases the antioxidant capacity of the cells as well as it can lead to the release of toxic ROS [21, 65], wellacceptable evidence of its two faces properties. Therefore, it has both properties in the antioxidant network and redoxregulated signaling in cells [66]. It has been demonstrated that labile Zn²⁺-coordination environments with cysteine ligands oxidizing the sulfur-ligands together with reducing with concomitant release and binding of labile Zn^{2+} [45, 53, 65, 67]. Moreover, early studies have been demonstrated that high intracellular labile Zn²⁺ elevates ROS in living cells by activating the enzyme nicotinamide adenine dinucleotide phosphate (NADPH) oxidase [67, 68]. Besides, in another study, it has been shown that labile Zn²⁺ can protect cells against oxidative damage through acting on the stabilization of membranes and inhibiting NADPH-oxidase, which is a pro-oxidant enzyme and induces metallothionein synthesis [69–71]. Besides, other studies mentioned that it can act as an antioxidant by affecting the expression of glutamatecysteine ligase to neutralize free radicals directly or indirectly [72-74]. Under hyperglycemic conditions, such as diabetes, studies demonstrated zinc-associated improvements in insulin sensitivity and glycemic control through reduction of the synthesis of ROS, thereby inhibiting the activation of oxidative stress pathways [75]. Those studies emphasized a zinc-favorous action on glucose transport into the cells [76, 77]. Together, hyperglycemic cardiomyocytes had high basal labile Zn²⁺, being associated with increased levels of not only increased ROS but also increased RNS in those cardiomyocytes [28, 78]. Furthermore, we have demonstrated that an antioxidant application could provide a balanced oxidant/ antioxidant level in the heart due to the prevention of the altered cellular redox state, though directly normalization of macromolecular complex responsible for both intracellular Ca²⁺- and Zn²⁺-homeostasis in hyperglycemic cardiomyocytes from the diabetic rats [25]. Studies emphasized how it is important to maintain an adequate concentration of zinc in the cell compartments for the essentiality of the proper functioning of the antioxidant defense system. Moreover, oxidative stress appears to be capable of altering the expression of proteins responsible for the Zn²⁺-homeostasis [79].

The ion Zn²⁺ can act as a pro-oxidant when its concentration is either deficient or in excess and becomes pro-inflammatory and pro-apoptotic, whereas it has an important role in the antioxidant defense system through regulation of glutathione peroxidase and in the expression of metallothionein, as well as it is a co-factor for superoxide dismutase. Interestingly, it has been also shown that a low zinc concentration could induce an important level of oxidative stress which further leads to cell death and promotes the production of ROS [80, 81]. It is noteworthy that, zinc as a multifunctional micronutrient, intracellular labile Zn²⁺ in biological systems has two faces, particularly under pathophysiological conditions, at most, depends on its level.

Labile Zn²⁺-mediated alterations in cardiomyocytes through its phosphorylation and oxidation actions of intracellular proteins

Several in vivo and in vitro studies strongly indicate that systemic and cellular Zn^{2+} -homeostasis are important processes in mammalian life and are controlled with different regulatory proteins. Intracellular labile Zn^{2+} in cardiomyocytes has multiple functions to provide cardioprotection in the preventions of different pathological conditions in the



heart. Although zinc is important against oxidative stress and cytoprotection processes in the heart, its role in induction together with regulation of proteins remains largely not known yet. Correspondingly, we have shown that hyperglycemic cardiomyocytes from experimental diabetic rats have higher resting intracellular labile Zn²⁺ level, linking increased both ROS and RNS levels in those cardiomyocytes [25, 28, 57]. In further observations, we determined a marked decrease in the activity of protein phosphatase 1 and 2A, a significant increase in the phosphorylation levels of extracellular signal-regulated kinase1/2, RyR2, and accessory protein of RyR2 macromolecular complex, FKBP12.6, as well as protein kinase A (PKA) and calcium calmodulin kinase II (CaMKII). To confirm the high intracellular labile Zn²⁺ induced changes in those proteins and kinases, we performed in vitro studies with rat ventricular cardiomyocytes incubated with either a zinc-ionophore of 1-hydroxy pyridine-2-thione or ZnCl₂. Then we determined first the phosphorylation levels of RyR2 and FKBP12.6 and then the phosphorylation levels of PKA and CaMKII together with activation in transcription factors such as NFkB and GSK and other endogenous actors such as Akt [25, 26]. There were marked increases in the phosphorylation levels of those proteins and kinases in those incubated cells. In early studies, we have also demonstrated that either high labile Zn²⁺ or increased oxidative stress could induced markedly increased levels of oxidation in protein thiols [21, 45, 66, 82]. Further studies supported our above results. They have shown that high intracellular labile Zn²⁺ inhibits the activity of adenylyl cyclases, the hormone, and forskolin stimulation of cAMP synthesis in N18TG2 cells [83]. It also caused inhibition of substrate phosphorylation by CaMKII such as to produce a concentration-dependent inhibition of phospholamban phosphorylation in the presence of Ca²⁺ and calmodulin [84]. Those above observations, under in vivo and in vitro high Zn²⁺ conditions, further supported the hypothesis that a Zn²⁺-disbalance could affect different signaling pathways resulting in several cellulars in different signaling networks. Among them, the critical roles of intracellular high labile Zn²⁺ in the redox signaling pathway together with its role in maintaining the normal structure and physiology of cellular actors should be one of the main reasons besides others [53, 85–90]. Supporting to those data, early studies mentioned that Zn²⁺ has multiple functional effects on kinases including PKC and cAMP-dependent protein kinase [91].

Overall, one can propose that intracellular high labile Zn^{2+} in cardiomyocytes under pathological conditions, seems to be closely associated with alterations in several cellular proteins, responsible for higher levels of phosphorylation and oxidation of the actors of this machinery as well as a high level of ROS and RNS. Therefore, it can be summarized that an intracellular labile Zn^{2+} level is modulated by the redox state of the cells (being associated with the

levels of both ROS and RNS [92]. Indeed, zinc-coordination environments with cysteine ligands have a property in which the sulfur-ligands can be oxidized and then reduced with concomitant release and binding of labile Zn²⁺ while it is about 30% buffering capacity emanates from sulfur donors (thiols), serving as redox buffer capacity [92, 93]. However, all the above effects strongly are depending on its level in cells. Zn²⁺ can increase the antioxidant capacity of the cells beside it can lead to the release of toxic ROS [19, 28, 45]. So far, the cellular toxicity of excess labile Zn²⁺ in cardiomyocytes can induce a dyshomeostasis in intracellular labile Ca²⁺, and thereby, an impairment in excitation-contraction coupling, as well as high-level production of ROS and/or RNS and loss of signaling quiescence leading to apoptosis in cells and cell death [19, 39, 45, 53, 54, 94, 95].

Zn²⁺-transporters mediate the control of cellular Zn²⁺ among intracellular compartments of cardiomyocytes

Together, our studies and literature data performed in mammalian tissues as well as human heart tissues provide strong evidence for two faces of zinc as a supplement or toxic through intracellular labile Zn²⁺ in the function of organs under physiological and pathological conditions, including diabetes, metabolic syndrome or obesity [18, 26, 28, 54, 96–100]. Correspondingly, studies have shown how low levels of zinc have adverse effects on physiological and metabolic functions (particularly linked to obesity) in humans as well as its high levels are detrimental to organs including the heart [18, 19, 28, 47, 54, 96]. Today, it is well documented that cellular homeostasis of labile Zn²⁺ is regulated and controlled efficiently with two families of specific Zn²⁺-transporters. One family named SLC39A family has 14 members and functions to carry labile Zn²⁺ into the cytosol in cells (ZIPs) whereas the second family is the SLC30A family which has 10 members and carries labile Zn²⁺ out off cytosol (ZnTs). Alterations in their expression and/or localization can lead to intracellular labile Zn²⁺ homeostasis which can underline several pathophysiological stimuli further leading to cellular damages [48, 57, 95, 101, 102]. Also, there is a close correlation between alterations in intracellular labile Zn²⁺ level and progression of many diseases including heart diseases, therefore, alterations in expression and/or function of any Zn²⁺-transporters can be one of the reasons for the development of diseases in mammalians. This event is a strong clue why those transporters are playing important roles in a human health situation.

ZIPs are expressed in different cell types in mammalians which regulate intracellular free Zn²⁺ and have crucial roles in physiology and pathophysiology. It is shown that ZIP1 [103–108], ZIP2 [107–110], ZIP3 [107–110], ZIP7 [57, 79,



111–116] and ZIP8 [79, 105, 115, 117–119] are identified in widespread mammary tissues and cells. Besides, ZIP4 protein is found in skin, chondrocytes, odontoblasts, fibroblast, pancreas, gastrointestinal tract, kidney, and hippocampal neurons [120–123], ZIP5 is found in the pancreas, kidney, liver, stomach, intestine, and hepatocytes [120–123], ZIP6 is found in several cancer tissues, neuroblastoma cells, T lymphocytes, peripheral blood mononuclear cells [124–130], while ZIP9 is found in the prostate, HeLa cells [131, 132]. ZIP10 has been shown in testis, kidney, breast, pancreatic α -cells [118, 119, 133–136], whereas ZIP11 is found in testis and digestive system, glands [110, 137, 138]. Further studies have shown that ZIP12 is found in the brain, lung, testis, and retina, neurons, endothelial, smooth muscle, and interstitial cells [110, 139, 140], while ZIP13 is found in bone, fat and adipose tissue, and also in hepatocytes [115, 141–143]. The last member of the ZIPs family, ZIP14 has been shown in bone and adipose tissue [79, 115, 135, 144-147]. The expressions of ZIP7, ZIP8, and ZIP14 have also been shown in hepatocytes and heart, as well [29, 148].

In mammalian tissues and cells, it has been identified 10 ZnTs in that member, which are responsible for Zn²⁺ efflux from the cytosol in cells. ZnTs are expressing in different types of tissues and cells including the brain, liver, gut, fat, heart, intestine, stomach, prostate, retina, pancreas, testis, muscle, and many types of cells including secretory cells and pancreatic β-cells. Studies demonstrated that ZnT1 presents in peripheral blood mononuclear cells [104–107, 130, 149, 150], whereas ZnT2 is found in the mammary gland, prostate, retina, pancreas, small intestine, and kidney [103–107, 110], ZnT3 is found in prostate glands [106, 107, 109, 110, 151], while ZnT4 is found in various tissues such as skin, chondrocytes, odontoblasts and fibroblast, pancreas, gastrointestinal tract, kidney, and hippocampal neurons [120–123, 141], ZnT5 is found in bone and heart [79, 105, 123, 152, 153]. ZnT6 is generally found in cancer tissues, and neuroblastoma cells, T lymphocytes, peripheral blood mononuclear cells [124-126, 128-130]. ZnT7 is found in different main organ tissues such as the brain, liver, gut, fat, heart, intestine, stomach, prostate, retina, pancreas, testis, muscle, and many types of cells including secretory cells, pancreatic β-cells [29, 48, 57, 111, 112, 116, 154–159]. ZnT8 is found in the pancreas, thyroid, heart, testis, and several cell types including cardiomyocytes, islet cells, pancreatic cells, endocrine cells, adrenal glands, insulin granules, pancreas, thyroid, adrenal gland [48, 57, 159-170]. The last two members of that family, ZnT9 is found in prostate, brain, muscle, kidney, HeLa cells [131, 171, 172], while ZnT10 is found in testis, kidney, breast, pancreatic α -cells, red blood cells, brain, liver, erythroid, and kidney [118, 119, 133–135, 173, 174].

Labile Zn²⁺ is not only an essential structural constituent of many intracellular actors but also it has a central role in

excitation-contraction coupling in cardiomyocytes. Therefore, any change in its physiological range could initiate induction of deleterious changes directly and/or indirectly in the heart [19, 45, 53]. In those considerations particularly in recent years, there are some research and review articles mentioned why Zn^{2+} -transporters are important for several organ proper functions in mammalians through being responsible for the re-distribution of subcellular labile Zn^{2+} levels at cell levels. For instance, in the last 5 years, it is published over 200 articles focused on the impact of Zn^{2+} -transporters in health and disease [47, 48, 102, 175–188].

The already shown roles of already known several Zn²⁺-transporters (for sure not all) are summarized in Tables 1 and 2 with their references. The phenotypes of those Zn²⁺-transporters knockout mice and variants have been also characterized in mammalian tissues and cells [117, 189–191] and the results of early studies on Zn²⁺-transporters are under consideration particularly during the last 20 years [106, 110, 177, 179–181, 183, 192–198].

Structure and function of mitochondria in cardiomyocytes under pathophysiological conditions via high intracellular labile Zn²⁺

Mitochondria in the mammalian heart are the major sources of the high-energy compound, ATP, which have multiple activities, and one of the vital organelles in eukaryotes including cardiac cells, as well [2, 6, 218]. Mitochondria are classified as either subsarcolemmal or interfibrillar in cardiomyocytes. There are two aqueous spaces such as the intermembrane space and the matrix of two lipid bilayer membranes, while the outer membrane has a role as the boundary between the cytoplasm and mitochondria. Importantly, that part contains multiple receptors and transporters to perform communication between mitochondria and other organelles, such as Sarco(endo)plasmic reticulum, SER, as well as cytoplasm [171, 219-221]. The morphology of cardiac mitochondria, as well as their physiology, is available to support the cell viability under different pathological situations, such as diabetes or aging [25, 27, 29]. Correspondingly, studies emphasize a close apposition between SER and mitochondria representing a key platform responsible for the regulation of different fundamental cellular pathways under physiological conditions, including redox-regulation of the cells [222]. Studies imply that any alteration in the SER-mitochondria axis can cause an onset and progression of several diseases, including cardiovascular disorders [29, 48, 223, 224].

Mitochondria play a central role in the heart homeostasis in mammalians. In general, electron microscopy of analysis of cardiac mitochondria showed that they have an



Table 1 Distribution of Zn^{2+} -transporters in mammalian tissues/cells responsible of Zn^{2+} -influx into cytosol (ZIPs)

Names of proteins	Types of tissues/Cells	References	
ZIP1	Widespread mammary tissues and cells	[103–108]	
ZIP2	Widespread mammary tissues and cells	[103–105, 107, 108]	
ZIP3	Widespread mammary tissues and cells, prostate glands	[107–110]	
ZIP4	Skin, chondrocytes, odontoblasts and fibroblast, pancreas, gastrointestinal tract, kidney, and hippocampal neurons	[120–123]	
ZIP5	Pancreas, kidney, liver, stomach, and intestine, hepatocytes	[79, 105, 153, 264]	
ZIP6	several cancer tissues, neuroblastoma cells, T lymphocytes, peripheral blood mononuclear cells	[124–130]	
ZIP7	Widespread mammary tissues and cells, hepatocytes, cardiomyocytes	[57, 79, 111–116]	
ZIP8	Widespread mammary tissues and hepatocytes, red blood cells,	[79, 105, 115, 117–119]	
ZIP9	Prostate, HeLa cells	[131, 132]	
ZIP10	Testis, kidney, breast, pancreatic α cells, red blood cells, brain, liver, erythroid, and kidney	[118, 119, 133–136]	
ZIP11	Testis and digestive system, glands	[110, 137, 138]	
ZIP12	Brain, lung, testis, and retina, neurons, endothelial, smooth muscle, and interstitial cells	[110, 139, 140]	
ZIP13	Bone, fat tissue, adipose tissue, hepatocytes	[115, 141–143]	
ZIP14	Bone, adipose tissue, bone, liver, heart, placenta, lung, brain, pancreatic α -cells	[79, 115, 135, 144–147]	

elliptical shape with either lamelliform or tubular numerous transverse cristae. They have also numerous sharp angulations, mall dense granules which are deposits of divalent cations present in the mitochondrial matrix [225]. The Zn²⁺ is required in the matrix of the mitochondria for the function of proteins and special ion transporters within mitochondrial compartments [226–232]. Labile Zn²⁺ is detected in the mitochondria of mammalian neuronal cells [231], which is compartmentalized into the mitochondrial membrane [231] associated with release from that compartment further leading to cell death [229].

It can be stated that labile Zn²⁺ can be detected in the mitochondria of mammalian cardiac cells using Zn²⁺-responsive fluorophores [47, 50, 230] [. Although the mitochondrial labile Zn²⁺ is low compared to either cytosol or SER in cardiomyocytes under physiological conditions, it can increase over normal values under pathological conditions, including hyperglycemia [47]. Even early studies mentioned the toxic effects of elevated intracellular labile Zn²⁺ for mammalian cells through its action on the modulation of gene expression and mitochondrial function [43, 45, 233, 234]. Furthermore, it has been pointed out the importance of an optimal range for the ratio of intracellular Zn²⁺ to Ca²⁺ in both cytosol and mitochondria to protect cardiomyocytes via controlling oxidative stress through regulation of mitochondrial function with Zn²⁺ [46, 235]. Additional studies have also shown a close association between elevated cytosolic labile Zn²⁺ and impairment of mitochondrial respiration under pathological stimuli in mammalian cells [235, 236].

Some studies indicate that there is a close relation between mitochondrial Zn²⁺ and mitochondrial membrane potential in either neurons or cardiomyocytes [28, 47, 228, 230]. It is an interesting process that any disruption of

mitochondrial membrane potential results in the release of Zn²⁺ to the cytosol whereas high labile Zn²⁺ can induce serious disruption of mitochondrial membrane potential in those cells. This release of mitochondrial labile Zn²⁺ can be a contributing cause of cellular damage and/or death during pathological stimuli [28, 229]. Interestingly, Dineley and co-workers [237] have shown a loss of membrane potential and elevation of ROS in rat brain mitochondria by high Zn²⁺. One of the impacts of combined effects of labile Zn²⁺ and Ca²⁺ is on the openings of mitochondrial permeability transition pore and increased the production of ROS, which are also closely associated with the induction of ER stress and apoptosis [238, 239]. Likely, the mitochondrial membrane potential is known to be not only an important driving force for ATP production during oxidative phosphorylation, but also for the mitophagy, and for the transport of proteins and ions such as Ca²⁺ and Zn²⁺ in cells including cardiomyocytes [10, 18, 29, 48, 240].

Zinc is generally as Zn²⁺ in biological macromolecules of mammalian cells [31, 36, 38, 39], however, it can be very toxic to most living cells when they expose to it beyond its normal physiological levels [28, 45, 241]. Being one of the most affected organelles, mitochondria in cardiomyocytes have detectable labile Zn²⁺ besides labile Ca²⁺ [27, 29, 48]. Although mitochondrial labile Zn²⁺ level is low compared to the cytosol and SER in cardiomyocytes it can get very high under pathological conditions, such as hyperglycemia and hyperinsulinemia as well as aging [10, 47, 48, 50]. Exposure to high Zn²⁺ and/or increases in intracellular labile Zn²⁺ via different signaling stimuli can increase the mitochondrial labile Zn²⁺ level while it, in turn, induces serious increases in ROS production and decreases in ATP level of cardiomyocytes [18, 47, 48]. More importantly, we, here and



Table 2 Distribution of Zn²⁺-transporters in mammalian tissues/cells responsible for Zn²⁺-efflux of cytosol (ZnTs)

[104–107, 130, 149, 150]
5100 10F
If one wants to give challenging examples on Zn ²⁺ -transporters it will include the involvement of ZnT1, ZIP4, and ZlP5 in intestinal zinc-transport, the involvement of ZlP10 and ZnT1 in renal zinc-reabsorption, and the roles of ZlP5, ZnT2, and ZnT1 in the pancreatic release of endogenous-zinc in the handling of dietary-zinc [193]. Further studies demonstrated the major factors in the regulation of Zn ²⁺ -homeostasis such as theinvolvement of ZnT2 in lactation, ZIP14 in the hypozincemia of inflammation, ZIP6, ZIP7, and ZIP10 in metastatic breast cancer, and ZnT8 in insulin processing and diabetes [177, 179–181, 183, 196–198]. Moreover, Ellis et al. [199] demonstrated the important contribution of a cytosolic Zn ²⁺ -importer transporter, ZIP7 in releasing Zn ²⁺ from the S(E)R, However, Huang et al. [111] showed the ZIP7 localization to the Golgi apparatus in CHO cells, while others demonstrated the roles of ZIP7 in the facilitation of Zn ²⁺ release of from the ER and behaves as a critical component in the subcellular re-distribution of Zn ²⁺ in cancer cells [200, 201]. Besides, ZnT7 was shown as a novel mammalian Zn ²⁺ -transporter, accumulates Zn ²⁺ in the Golgi apparatus as well as into cytosol from S(E) R and mitochondria [29, 112, 202]. There are important data showed why changes in the expression and activity of different Zn ²⁺ -transporters have been directly linked to both systemic and organ level diseases, as well as rare diseases such as acrodermatitis enteropathica [114, 120, 124, 203, 204]. One group of highlighted studies on the role of Zn ²⁺ -transporters in health and disease includes the studies in the nervous system, including the role of high cytosolic Zn ²⁺ and ZIP12 in neuronal differentiation [139]. Similar to the above studies, it has been documented that ZnT3 is critical for the transport of Zn ²⁺ into synaptic vesicles of a subset of glutamatergic neurons [205], and its expression is reduced in patients with Alzheimer's disease [206] and Parkinson; disease-related dementia [207]. H

previously, have shown that exposure to high Zn²⁺ induced marked increases in mitochondrial matrix/cristae area and matrix volume together with an increased lysosome in cardiomyocytes [26, 179]. Together, the notable clustering and vacuolated mitochondrion markedly disrupted and damaged myofibrils, and electron-dense small granules were observed in Zn²⁺-exposed cardiomyocytes [26]. Those changes were also including notable increases in mitochondrial matrix/cristae area and matrix volume, together with some signs indicating fission-fusion defects in the mitochondria, in a manner of its concentration-dependent [26]. High Zn²⁺ exposure also caused a marked depolarization in mitochondrial membrane potential, as well [28, 29, 48]. Additional

studies have also shown a close association between intracellular high labile Zn^{2+} and impairment of mitochondrial respiration in a variety of pathological conditions in mammalian cells [235, 236]. One can state that if intracellular labile Zn^{2+} gets over its physiological level, it can stimulate one or more deleterious changes, such as marked alterations in mitochondrion morphology and function as well as marked changes in the phosphorylation/oxidation levels of cytosolic signaling proteins [47, 48]. Moreover, it has been demonstrated that both extra-and intracellular high-level Zn^{2+} modulates L-type Ca^{2+} -channel properties, as well as its regulation by β -adrenergic agonists independently of altering the cellular redox status but associated with cellular



ATP level [93]. However, in an early study by Traynelis et al. demonstrated contradictory data demonstrating the inhibition of both L-type and T-type Ca^{2+} currents with high Zn^{2+} in neuronal cells [242]. Correspondingly, others had demonstrated a more sensitivity of K^+ -channels to high Zn^{2+} than those of Na^+ -channels in neural cells [243], whereas a recent data has been shown activation of the M-type (including Kv7 channels) K^+ -channels by high intracellular labile Zn^{2+} [244].

Here, we incubated ventricular cardiomyocytes with different zinc-compounds and using light and electron microscopy analysis, the heart tissue, and cardiomyocytes. The electron microscopy analysis showed that incubation of cardiomyocytes with a Zn²⁺-ionophore, Zn²⁺-pyrithione (ZnPT; 0.01-µM for 1-h) induced elongation in mitochondria leading to a significant increase in a sarcomere length, and clear irregular cristae appearance of mitochondrion located between myofibrils, together with electron-dense matrix (Fig. 1A, left). A tenfold increase in ZnPT concentration induced marked changes in the shapes of the mitochondria such as fragmentation, rounding, and swollen (Fig. 1A, middle). In incubation of the cells with the highest ZnPT concentration (1-µM), the mitochondria appeared more electron-lucent while the loss of the matrix density (Fig. 1A, right). When cardiomyocytes incubated with 10-μM ZnPO₄ (1-h), there was more disorganized mitochondrial cristae, and electron-lucent matrix, and partitioned mitochondria in the cells (Fig. 1B, left). The cardiomyocytes incubated with 0.1 µM ZnCl₂ (1-h), clustered mitochondria, slight intramitochondrial edema, and enlargement of T-tubules and highly localized lysosomes were observed (Fig. 1B, right). In this regard, it has been demonstrated concentration-dependent Zn²⁺ inhibition of mitochondrial complex I [236], as well as Zn²⁺ entry into mitochondria via uniporter inducing mitochondrial dysfunction, at most, via ROS production and contributing to mitochondrial Ca²⁺ deregulation [245].

As a consequence mentioned above paragraphs, the impaired mitochondrial function through exposure to high Zn²⁺ and/or increase intracellular labile Zn²⁺ might lead to several cardiovascular diseases. Therefore, one can emphasize the importance of a well-controlled intracellular labile Zn²⁺ through the mitochondria as a novel therapeutic target for cardiac complications under pathological conditions including oxidative stress. Indeed, studies pointed out that cardiac mitochondria, similar to SER, also play an important role in regulating not only Ca²⁺-homeostasis but also Ca²⁺-homeostasis via acting as a sponge to buffer both ions in cardiomyocytes [19, 21, 25, 29, 45, 48]. So far, it has

been shown that both elevated labile ion levels such as Zn^{2+} and Ca^{2+} in the cytosol are deleterious in cardiomyocytes, and therefore their well-controlled levels in the cytosol are necessary to maintain a physiologic function of the heart. Supporting the last statement, we, recently, have shown that mitochondria played an important role to maintain cytosolic labile Zn^{2+} level though uptake high Zn^{2+} from cytosol increased due to high-level release from SER in hyperglycemic or hypertrophic ventricular cardiomyocytes [29, 48]. Therefore, one can interpret that mitochondria contribute to cellular Zn^{2+} -muffling between cellular compartments under pathological conditions via affecting S(E)R-mitochondria coupling [246–250].

Distribution of Zn²⁺-transporters in mitochondria of cardiomyocytes

Similar to others, there are several Zn²⁺-signaling pathways to control the intracellular Zn2+ homeostasis in cardiomyocytes. Of note, the intracellular Zn²⁺-signaling can easily interfere with the Ca²⁺-signaling in cardiomyocytes, under both physiological and pathological conditions [19–21, 25, 45, 58]. A piece of widespread information on cellular regulation of cytosolic Zn²⁺-signaling through Zn²⁺-transporters, Zn²⁺-binding molecules, –fingers, and Zn²⁺-sensors in several tissues and cell types are very well documented [96, 111, 190, 193, 199, 200, 251–255], the distribution and function of those carries in subcellular organelles are not well clarified in cardiomyocytes yet.

Recently we and others have demonstrated that Zn²⁺-transporters induced developmental and physiological defects in mammalians including cardiomyopathy in the heart [27, 29, 57]. Following demonstrating the distribution of labile in the cytosol, SER, and mitochondria of cardiomyocytes using eCALWY probes [50] and the important roles of both ZIP7 and ZnT7 to mediate ER stress in hyperglycemic cardiomyocytes [57], we first demonstrated the subcellular localizations of ZIP8, ZIP14 and ZnT8 in cardiomyocytes besides ZIP7 and ZnT7 in cardiomyocytes [148]. By using the Huygens program for co-localization values of those transporters, we calculated Pearson's coefficients (PC) for ZIP8-SER and ZIP8-sarcolemma as $44 \pm 3\%$ and $60 \pm 2\%$, respectively. The PC values of ZIP14 were $50 \pm 8\%$ and $42 \pm 3\%$ for SER and sarcolemma, while those PC values of ZnT8 were $66 \pm 3\%$ and $80 \pm 2\%$ for SER and sarcolemma [148]. Those PCs strongly supported the high-level localization of those three Zn²⁺-transporters on sarcolemma ventricular



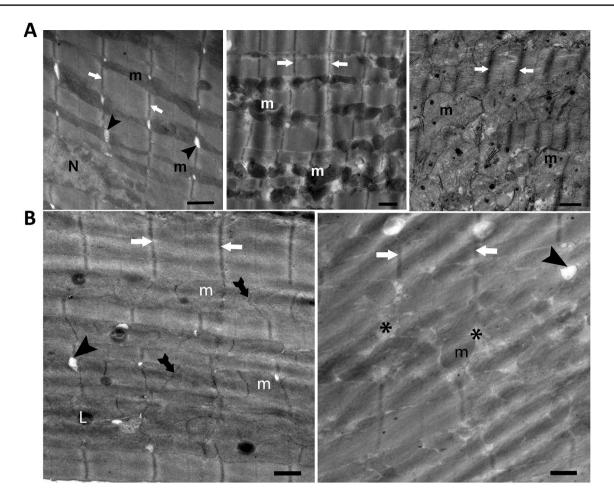


Fig. 1 The electron microscopy analysis of left ventricular cardiomyocytes incubated with a Zn²⁺-ionophore, Zn²⁺-pyrithione, ZnPT (0.01-μM, 0.1-μM, or 1-μM for 1-h) (A; left, middle, right, respectively), with 10-μM ZnPO₄ (1-h; B, left), or with 0.1 μM ZnCl₂ (1-h;

B, right). Shorten symbols; m: mitochondria, arrow: Z-line, L: lysosome, N: nucleus, tailed arrow: partitioned mitochondrion, arrowhead: T-tubule, asterisk: intramitochondrial edema. Magnification: ×12,930 and bars: 500 nm

cardiomyocytes. In the same study, authors demonstrated that the expression levels of ZIP14 and ZnT8 were significantly high in the human heart with serious failure, whereas ZIP8 level was significantly low than those of controls, through, at most, increased oxidative and ER stress. Correspondingly, we have shown that the expression levels of ZIP7, ZnT7, and ZIP14 were decreased with no change in ZIP8 of high carbohydrate diet-induced metabolic syndrome rat cardiomyocytes [102]. Furthermore, in our other study, there were significant increases in the expression levels of ZIP7, ZIP14, and ZnT8 along with decreases in the ZIP8 and ZnT7 levels in the heart tissue from transverse aortic constriction model induced hypertrophic young rats [159, 202].

Recently, authors also studied the role and localization of Zn^{2+} -transporters on mitochondria in aged ventricular cardiomyocytes. Together with high ROS level in those cells, the examination of the distribution of cellular labile Zn^{2+} among suborganelles, such as S(E)R and mitochondria parallel to cytosolic labile Zn^{2+} showed that the cytosolic was markedly high, at most, due to increased ZIP7 level with decreased ZnT7 level [48]. In that study, it was for the first time demonstrated that labile Zn^{2+} level in isolated mitochondria was significantly high while it was decreased in isolated SER, supporting the hypothesis of re-distribution of Zn^{2+} -transporters under the pathological condition to buffer the intracellular labile Zn^{2+} level.



Supporting the re-distribution of labile Zn²⁺ among cytosol and organelles through Zn²⁺-transporters, the Western-blotting data demonstrated that the levels of ZnT7 and ZnT8 were increased in isolated mitochondria with no changes in ZIP7 and ZIP8 levels [48]. Those changes have positive responses to the mitochondria-targeting antioxidant (MitoTEMPO) treatment of those cells, as well. Moreover, another transporter, the ZIP14 protein level was significantly low in isolated mitochondria from aged ventricular cardiomyocytes with a positive response to an application of the mitochondria targeting antioxidant [256].

Correspondingly, early studies pointed out a relatively low expressing levels of Zn²⁺-transporters such as ZIP7 and ZnT7 in mammalian heart tissues [111, 112, 235]. An interesting study by Seo et al. focused on showing the localization of ZnT2 in mammary epithelial cells (HC11) and they found that ZnT2 localized to the inner mitochondrial membrane and acts as an auxiliary Zn²⁺ importer into mitochondria [257]. In a recent study, authors also have shown the localization of ZIP1 on mitochondria and responsible for Zn²⁺-entry into mitochondria in HeLa cells [258]. Although limited data are demonstrating the importance of mitochondrial labile Zn²⁺ and the mitochondrial localization of Zn²⁺-transporters, our and earlier studies emphasized the role of excess labile Zn²⁺ likeness to Ca²⁺, in the injury of cells, including cardiomyocytes, through excess ROS production alone and/or together with mitochondrial dysfunction [26, 28, 234, 259–261]. However, there are controversies about how high Zn²⁺ can affect mitochondria function: Excess Zn²⁺ could induced increases have been reported to induce mitochondrial Zn²⁺ uptake, resulting in a longer loss of mitochondrial membrane potential in cultured neurons, besides prolonged duration of ROS production [44], whereas other reports demonstrated that high-level Zn²⁺ did not acutely depolarize mitochondria [262, 263]. Besides, a high Zn²⁺ could induce a clear depolarization in mitochondrial membrane potential parallel to high ROS production ventricular cardiomyocytes while high intracellular Zn²⁺ including hyperglycemic ventricular cardiomyocytes presented high ROS production as well as a clear depolarized mitochondrial membrane potential [28, 29, 57]. All the above studies are calling an important question whether or not high labile Zn²⁺ is an effective inhibitor of mitochondrial function under any pathological stimuli, therefore, this event is providing an important interest to a clarification of that question.

The already known documents showing re-distribution of some Zn²⁺-transporters localized to the mitochondria in mammalian ventricular cardiomyocytes under pathological conditions are summarized in Table 3.

Table 3 The re-distribution of some Zn^{2+} -transporters localized to the mitochondria in mammalian ventricular cardiomyocytes under pathological conditions

Types of proteins	Hypergly- cemic heart	Hyper-insu- linemic heart	Aged	Dilated/ Ischaemic/
	cells	cells	cells	Hypertropic heart cells
ZIP7	\downarrow	\leftrightarrow	\leftrightarrow	\downarrow
ZIP8	\downarrow	\leftrightarrow	\leftrightarrow	\leftrightarrow
ZIP14	\leftrightarrow	\downarrow	\downarrow	\leftrightarrow
ZnT7	↑	↑	↑	↑
ZnT8	\uparrow	↑	↑	↑

Here, the symbols \uparrow , \downarrow , and \leftrightarrow are representing increased, decreased and unchanged protein expression levels in associated pathological conditions (re-organized from references, 29, 48, 57, 82, 148, 159, 179, 256). All measurements are performed in isolated ventricular rat cardiomyocytes. All changes are statistically significant compared to those of control cardiomyocytes (p < 0.05)

Conclusions

Considering the already shown data, it is acceptable to mention the intracellular labile Zn²⁺ as a critical signaling molecule in normal cell physiology as well as in pathophysiological conditions, such as aging, diabetes, insulin resistance, or heart failure in mammalians. As mentioned previously, cellular Zn²⁺-homeostasis is tightly controlled by different regulatory signaling pathways including Zn²⁺-transporters alone and/or the pathways associated with Zn²⁺-transporters. In another insight, coordinated regulation of Zn²⁺ uptake, efflux, distribution, and storage in cardiomyocytes is a very important issue for a proper heart function in humans. Although experimental data clearly show the multiple biologic functions of intracellular labile Zn²⁺ there are yet some controversies among them, and, therefore, none of them are more clear than the others to provide cardioprotection in pathological cardiac tissue. Overall, here, we tried to document the prevalence of important relationships between intracellular labile Zn²⁺ and Zn²⁺-transporters, particularly localized to mitochondria, under physiological as well as under any pathological stimuli such as hyperglycemia, hyperinsulinemia, cardiomyopathy, heart failure, or aging (Fig. 2). Therefore, we first emphasized the possibility of an association between intracellular labile Zn²⁺ and Zn²⁺-transporters in mitochondria as therapeutic targets in heart dysfunction. Second, we proposed the importance of possible new therapeutic agents particularly targeting mitochondrial Zn²⁺-transporters, potentiality control that relationship in cardiac cells.



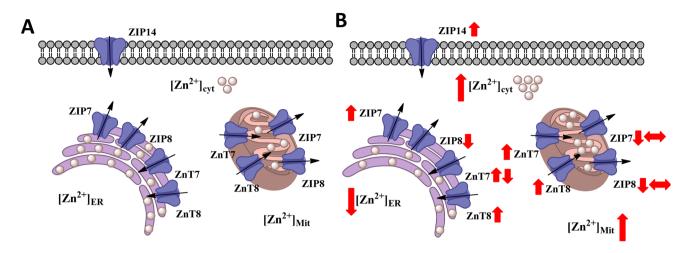


Fig. 2 A summarized representation to demonstrate the re-distribution of intracellular labile Zn^{2+} levels in the cytosol $([Zn^{2+}]_i)$, mitochondria $([Zn^{2+}]_{Mit})$, and Sarco(endo)plasmic reticulum $([Zn^{2+}]_{SER})$ as well as the Zn^{2+} –transporters in left ventricular cardiomyocytes

under any pathological stimuli (hyperglycemia, hyperinsulinemia, cardiomyopathy, heart failure, aging, etc.) (B) comparison to that of physiological condition (A). The presentation is summarized from our already published articles [29, 48, 57, 82, 148, 159, 179, 256]

Acknowledgments Thanks to Dr. D. Billur for her electron microscopy analysis. This work was supported by grants (No. SGAB-216S979) from The Scientific and Technological Research Council of Turkey.

Compliance with ethical standards

Conflict of interest The authors declare no conflicts of interest.

References

- Ernster L, Schatz G (1981) Mitochondria: a historical review. J Cell Biol 91:227s–255s. https://doi.org/10.1083/jcb.91.3.227s
- Yang D, Oyaizu Y, Oyaizu H, Olsen GJ, Woese CR (1985) Mitochondrial origins. Proc Natl Acad Sci U S A 82:4443

 –4447. https://doi.org/10.1073/pnas.82.13.4443
- Gray MW, Burger G, Cedergren R, Golding GB, Lemieux C, Sankoff D, Turmel M, Lang BF (1999) A genomics approach to mitochondrial evolution. Biol Bull 196:400–403. https://doi. org/10.2307/1542980
- Gustafsson AB, Gottlieb RA (2008) Recycle or die: the role of autophagy in cardioprotection. J Mol Cell Cardiol 44:654

 661. https://doi.org/10.1016/j.yjmcc.2008.01.010
- Hoppel CL, Tandler B, Fujioka H, Riva A (2009) Dynamic organization of mitochondria in human heart and in myocardial disease. Int J Biochem Cell Biol 41:1949–1956. https://doi. org/10.1016/j.biocel.2009.05.004
- Friedman JR, Nunnari J (2014) Mitochondrial form and function. Nature 505:335–343. https://doi.org/10.1038/nature12985
- Zhao Q, Sun Q, Zhou L, Liu K, Jiao K (2019) Complex regulation of mitochondrial function during cardiac development. J Am Heart Assoc 8:e012731. https://doi.org/10.1161/jaha.119.01273
- Bonora M, Wieckowsk MR, Chinopoulos C, Kepp O, Kroemer G, Galluzzi L, Pinton P (2015) Molecular mechanisms of cell death: central implication of ATP synthase in mitochondrial permeability transition. Oncogene 34:1608. https://doi.org/10.1038/ onc.2014.462

- Olgar Y, Tuncay E, Billur D, Durak A, Ozdemir S, Turan B (2020) Ticagrelor reverses the mitochondrial dysfunction through preventing accumulated autophagosomes-dependent apoptosis and ER stress in insulin-resistant H9c2 myocytes. Mol Cell Biochem 469:97–107. https://doi.org/10.1007/s11010-020-03731-9
- Olgar Y, Tuncay E, Degirmenci S, Billur D, Dhingra R, Kirshenbaum L, Turan B (2020) Ageing-associated increase in SGLT2 disrupts mitochondrial/sarcoplasmic reticulum Ca(2+) homeostasis and promotes cardiac dysfunction. J Cell Mol Med 24:8567–8578. https://doi.org/10.1111/jcmm.15483
- 11. Münzel T, Camici GG, Maack C, Bonetti NR, Fuster V, Kovacic JC (2017) Impact of oxidative stress on the heart and vasculature: part 2 of a 3-part series. J Am Coll Cardiol 70:212–229. https://doi.org/10.1016/j.jacc.2017.05.035
- Münzel T, Gori T, Bruno RM, Taddei S (2010) Is oxidative stress a therapeutic target in cardiovascular disease? Eur Heart J 31:2741–2748. https://doi.org/10.1093/eurheartj/ehq396
- Afanas'ev I (2011) ROS and RNS signaling in heart disorders: could antioxidant treatment be successful? Oxidative Med Cell Longev 2011:293769. https://doi.org/10.1155/2011/293769
- 14. Afanas'ev I (2011) Reactive oxygen species signaling in cancer: comparison with aging. Aging Dis 2:219–230
- Griendling KK, Touyz RM, Zweier JL, Dikalov S, Chilian W, Chen YR, Harrison DG, Bhatnagar A (2016) Measurement of reactive oxygen species, reactive nitrogen species, and redoxdependent signaling in the cardiovascular system: a scientific statement from the American Heart Association. Circ Res 119:e39–e75. https://doi.org/10.1161/res.00000000000000110
- Bray AW, Ballinger SW (2017) Mitochondrial DNA mutations and cardiovascular disease. Curr Opin Cardiol. https://doi.org/10.1097/hco.000000000000383
- Peoples JN, Saraf A, Ghazal N, Pham TT, Kwong JQ (2019) Mitochondrial dysfunction and oxidative stress in heart disease. Exp Mol Med 51:1–13. https://doi.org/10.1038/s12276-019-0355-7
- 18. Degirmenci S, Olgar Y, Durak A, Tuncay E, Turan B (2018) Cytosolic increased labile Zn(2+) contributes to arrhythmogenic action potentials in left ventricular cardiomyocytes through protein thiol oxidation and cellular ATP depletion. J



- Trace Elem Med Biol 48:202–212. https://doi.org/10.1016/j.jtemb.2018.04.014
- Tuncay E, Bilginoglu A, Sozmen NN, Zeydanli EN, Ugur M, Vassort G, Turan B (2011) Intracellular free zinc during cardiac excitation-contraction cycle: calcium and redox dependencies. Cardiovasc Res 89:634–642. https://doi.org/10.1093/cvr/cvq352
- Tuncay E, Okatan EN, Toy A, Turan B (2014) Enhancement of cellular antioxidant-defence preserves diastolic dysfunction via regulation of both diastolic Zn2+ and Ca2+ and prevention of RyR2-leak in hyperglycemic cardiomyocytes. Oxidative Med Cell Longev 2014:290381. https://doi.org/10.1155/2014/290381
- Ayaz M, Turan B (2006) Selenium prevents diabetes-induced alterations in [Zn2+]i and metallothionein level of rat heart via restoration of cell redox cycle. Am J Physiol Heart Circ Physiol 290:H1071–H1080. https://doi.org/10.1152/ajpheart.00754.2005
- Fukada T, Yamasaki S, Nishida K, Murakami M, Hirano T (2011) Zinc homeostasis and signaling in health and diseases: zinc signaling. J Biol Inorg Chem 16:1123–1134. https://doi.org/10.1007/s00775-011-0797-4
- Bellomo EA, Meur G, Rutter GA (2011) Glucose regulates free cytosolic Zn²⁺ concentration, Slc39 (ZiP), and metallothionein gene expression in primary pancreatic islet β-cells. J Biol Chem 286:25778–25789. https://doi.org/10.1074/jbc.M111.246082
- Woodier J, Rainbow RD, Stewart AJ, Pitt SJ (2015) Intracellular zinc modulates cardiac ryanodine receptor-mediated calcium release. J Biol Chem 290:17599–17610. https://doi.org/10.1074/jbc.M115.661280
- 25. Tuncay E, Okatan EN, Vassort G, Turan B (2013) β-blocker timolol prevents arrhythmogenic Ca²⁺ release and normalizes Ca²⁺ and Zn²⁺ dyshomeostasis in hyperglycemic rat heart. PLoS One 8:e71014. https://doi.org/10.1371/journal.pone.0071014
- Billur D, Tuncay E, Okatan EN, Olgar Y, Durak AT, Degirmenci S, Can B, Turan B (2016) Interplay between cytosolic free Zn(2+) and mitochondrion morphological changes in rat ventricular Cardiomyocytes. Biol Trace Elem Res 174:177–188. https://doi.org/10.1007/s12011-016-0704-5
- Olgar Y, Degirmenci S, Durak A, Billur D, Can B, Kayki-Mutlu G, Arioglu-Inan EE, Turan B (2018) Aging related functional and structural changes in the heart and aorta: MitoTEMPO improves aged-cardiovascular performance. Exp Gerontol 110:172–181. https://doi.org/10.1016/j.exger.2018.06.012
- Tuncay E, Turan B (2016) Intracellular Zn(2+) increase in Cardiomyocytes induces both electrical and mechanical dysfunction in heart via endogenous generation of reactive nitrogen species. Biol Trace Elem Res 169:294–302. https://doi.org/10.1007/s12011-015-0423-3
- Tuncay E, Bitirim CV, Olgar Y, Durak A, Rutter GA, Turan B (2019) Zn(2+)-transporters ZIP7 and ZnT7 play important role in progression of cardiac dysfunction via affecting sarco(endo) plasmic reticulum-mitochondria coupling in hyperglycemic cardiomyocytes. Mitochondrion 44:41–52. https://doi.org/10.1016/j.mito.2017.12.011
- Prasad AS (1981) Zinc deficiency and effects of zinc supplementation on sickle cell anemia subjects. Prog Clin Biol Res 55:99–122
- Vallee BL, Falchuk KH (1993) The biochemical basis of zinc physiology. Physiol Rev 73:79–118. https://doi.org/10.1152/ physrev.1993.73.1.79
- Sandstead HH, Prasad AS (2017) Dietary whole grains and zinc nutriture. Am J Clin Nutr 106:955–956. https://doi.org/10.3945/ ajcn.117.161224
- Bates CJ, Evans PH, Dardenne M, Prentice A, Lunn PG, Northrop-Clewes CA, Hoare S, Cole TJ, Horan SJ, Longman SC et al (1993) A trial of zinc supplementation in young rural Gambian children. Br J Nutr 69:243–255. https://doi.org/10.1079/bjn19 930026

- 34. Maxfield L, Crane JS (2020) Zinc deficiency. StatPearls Publishing Copyright © 2020, StatPearls Publishing LLC., Treasure Island (FL), StatPearls
- Caulfield LE and Black RE (2004) Zinc deficiency. Comparative quantification of health risks: global and regional burden of disease attributable to selected major risk factors 1:257–280
- Prasad AS (1985) Clinical, endocrinological and biochemical effects of zinc deficiency. Clin Endocrinol Metab 14:567–589. https://doi.org/10.1016/s0300-595x(85)80007-4
- Cousins RJ (1994) Metal elements and gene expression.
 Annu Rev Nutr 14:449–469. https://doi.org/10.1146/annurev.nu.14.070194.002313
- 38. Maret W (2011) Metals on the move: zinc ions in cellular regulation and in the coordination dynamics of zinc proteins. Biometals 24:411–418. https://doi.org/10.1007/s10534-010-9406-1
- Maret W (2013) Zinc and human disease. Met Ions Life Sci 13:389–414. https://doi.org/10.1007/978-94-007-7500-8_12
- Prasad AS (2014) Impact of the discovery of human zinc deficiency on health. J Trace Elem Med Biol 28:357–363. https:// doi.org/10.1016/j.jtemb.2014.09.002
- Prasad AS, Bao B (2019) Molecular mechanisms of zinc as a pro-antioxidant mediator: clinical therapeutic implications. Antioxidants (Basel) 8. https://doi.org/10.3390/antiox8060164
- Marreiro DD, Cruz KJ, Morais JB, Beserra JB, Severo JS, de Oliveira AR (2017) Zinc and oxidative stress: current mechanisms. Antioxidants (Basel) 6. https://doi.org/10.3390/antiox6020024
- Manev H, Kharlamov E, Uz T, Mason RP, Cagnoli CM (1997) Characterization of zinc-induced neuronal death in primary cultures of rat cerebellar granule cells. Exp Neurol 146:171–178. https://doi.org/10.1006/exnr.1997.6510
- 44. Sensi SL, Yin HZ, Weiss JH (1999) Glutamate triggers preferential Zn2+ flux through Ca2+ permeable AMPA channels and consequent ROS production. Neuroreport 10:1723–1727. https://doi.org/10.1097/00001756-199906030-00018
- Turan B, Fliss H, Désilets M (1997) Oxidants increase intracellular free Zn2+ concentration in rabbit ventricular myocytes.
 Am J Phys 272:H2095–H2106. https://doi.org/10.1152/ajpheart.1997.272.5.H2095
- 46. Chanoit G, Lee S, Xi J, Zhu M, McIntosh RA, Mueller RA, Norfleet EA, Xu Z (2008) Exogenous zinc protects cardiac cells from reperfusion injury by targeting mitochondrial permeability transition pore through inactivation of glycogen synthase kinase-3beta. Am J Physiol Heart Circ Physiol 295:H1227–h1233. https://doi.org/10.1152/ajpheart.00610.2008
- 47. Tuncay E, Olgar Y, Durak A, Degirmenci S, Bitirim CV, Turan B (2019) beta3 -adrenergic receptor activation plays an important role in the depressed myocardial contractility via both elevated levels of cellular free Zn(2+) and reactive nitrogen species. J Cell Physiol 234:13370–13386. https://doi.org/10.1002/jcp.28015
- 48. Olgar Y, Tuncay E, Turan B (2019) Mitochondria-targeting antioxidant provides Cardioprotection through regulation of cytosolic and mitochondrial Zn(2+) levels with re-distribution of Zn(2+)-transporters in aged rat Cardiomyocytes. Int J Mol Sci 20. https://doi.org/10.3390/ijms20153783
- Zhang K, Yang W, Zhang M, Sun Y, Zhang T, Liu J, Zhang J (2019) Pretreatment with antiplatelet drugs improves the cardiac function after myocardial infarction without reperfusion in a mouse model. Cardiol J. https://doi.org/10.5603/CJ.a2019.0051
- Chabosseau P, Tuncay E, Meur G, Bellomo EA, Hessels A, Hughes S, Johnson PR, Bugliani M, Marchetti P, Turan B, Lyon AR, Merkx M, Rutter GA (2014) Mitochondrial and ER-targeted eCALWY probes reveal high levels of free Zn2+. ACS Chem Biol 9:2111–2120. https://doi.org/10.1021/cb5004064
- Kamalov G, Deshmukh PA, Baburyan NY, Gandhi MS, Johnson PL, Ahokas RA, Bhattacharya SK, Sun Y, Gerling IC, Weber KT



- (2009) Coupled calcium and zinc dyshomeostasis and oxidative stress in cardiac myocytes and mitochondria of rats with chronic aldosteronism. J Cardiovasc Pharmacol 53:414–423. https://doi.org/10.1097/FJC.0b013e3181a15e77
- 52. Coudray C, Charlon V, De Leiris J, Favier A (1993) Effect of zinc deficiency on lipid peroxidation status and infarct size in rat hearts. Int J Cardiol 41:109–113
- Atar D, Backx PH, Appel MM, Gao WD, Marban E (1995) Excitation-transcription coupling mediated by zinc influx through voltage-dependent calcium channels. J Biol Chem 270:2473–2477
- Cicek FA, Toy A, Tuncay E, Can B, Turan B (2014) Beta-blocker timolol alleviates hyperglycemia-induced cardiac damage via inhibition of endoplasmic reticulum stress. J Bioenerg Biomembr 46:377–387
- 55. Murakami M, Hirano T (2008) Intracellular zinc homeostasis and zinc signaling. Cancer Sci 99:1515–1522
- Fukada T, Kambe T (2011) Molecular and genetic features of zinc transporters in physiology and pathogenesis. Metallomics 3:662–674
- 57. Tuncay E, Bitirim VC, Durak A, Carrat GR, Taylor KM, Rutter GA, Turan B (2017) Hyperglycemia-induced changes in ZIP7 and ZnT7 expression cause Zn2+ release from the sarco (endo) plasmic reticulum and mediate ER stress in the heart. Diabetes 66:1346–1358
- Tuncay E, Cicek FA, Toy A, Turan B (2014) Intracellular free zinc ion increase triggers hyperglycemia-induced cardiomyocyte dysfunction through endoplasmic reticulum stress. Biophys J 106:113a
- Truong-Tran AQ, Carter J, Ruffin RE, Zalewski PD (2001)
 The role of zinc in caspase activation and apoptotic cell death.
 Zinc Biochemistry, Physiology, and Homeostasis, Springer, pp 129–144
- Miao X, Sun W, Miao L, Fu Y, Wang Y, Su G, Liu Q (2013) Zinc and diabetic retinopathy. J Diabetes Res 2013
- Wong SH, Zhao Y, Schoene NW, Han CT, Shih RS, Lei KY (2007) Zinc deficiency depresses p21 gene expression: inhibition of cell cycle progression is independent of the decrease in p21 protein level in HepG2 cells. Am J Physiol Cell Physiol 292:C2175–C2184. https://doi.org/10.1152/ajpcell.00256.2006
- Malaiyandi LM, Dineley KE, Reynolds IJ (2004) Divergent consequences arise from metallothionein overexpression in astrocytes: zinc buffering and oxidant-induced zinc release. Glia 45:346–353
- Yaras N, Ugur M, Ozdemir S, Gurdal H, Purali N, Lacampagne A, Vassort G, Turan B (2005) Effects of diabetes on ryanodine receptor ca release channel (RyR2) and Ca2+ homeostasis in rat heart. Diabetes 54:3082–3088. https://doi.org/10.2337/diabetes.54.11.3082
- Maret W (2000) The function of zinc metallothionein: a link between cellular zinc and redox state. J Nutr 130:1455S–1458S
- Maret W (2009) Molecular aspects of human cellular zinc homeostasis: redox control of zinc potentials and zinc signals. Biometals 22:149–157
- Ayaz M, Ozdemir S, Yaras N, Vassort G, Turan B (2005) Selenium-induced alterations in ionic currents of rat cardiomyocytes. Biochem Biophys Res Commun 327:163–173
- Noh KM, Kim YH, Koh JY (1999) Mediation by membrane protein kinase C of zinc-induced oxidative neuronal injury in mouse cortical cultures. J Neurochem 72:1609–1616
- Pierce GN, Dhalla NS (1985) Mechanisms of the defect in cardiac myofibrillar function during diabetes. Am J Physiol-Endocrinol Metabolism 248:E170–E175
- Maret W and Krężel A (2007) Cellular zinc and redox buffering capacity of metallothionein/thionein in health and disease. Molecular medicine, BioMed Central, pp 371–375

- Chasapis CT, Loutsidou AC, Spiliopoulou CA, Stefanidou ME (2012) Zinc and human health: an update. Arch Toxicol 86:521–534
- Ruz M, Carrasco F, Rojas P, Codoceo J, Inostroza J, Basfi-Fer K, Valencia A, Vásquez K, Galgani J, Pérez A (2013) Zinc as a potential coadjuvant in therapy for type 2 diabetes. Food Nutr Bull 34:215–221
- Foster M, Samman S (2010) Zinc and redox signaling: perturbations associated with cardiovascular disease and diabetes mellitus. Antioxid Redox Signal 13:1549–1573
- Eide DJ (2011) The oxidative stress of zinc deficiency. Metallomics 3:1124–1129
- 74. Liang T, Zhang Q, Sun W, Xin Y, Zhang Z, Tan Y, Zhou S, Zhang C, Cai L, Lu X (2015) Zinc treatment prevents type 1 diabetes-induced hepatic oxidative damage, endoplasmic reticulum stress, and cell death, and even prevents possible steatohepatitis in the OVE26 mouse model: important role of metallothionein. Toxicol Lett 233:114–124
- 75. Lima VBDS, Sampaio FDA, Bezerra DLC, Moita Neto JM, Marreiro DDN (2011) Parameters of glycemic control and their relationship with zinc concentrations in blood and with superoxide dismutase enzyme activity in type 2 diabetes patients. Arquivos Brasileiros de Endocrinologia & Metabologia 55:701–707
- Vardatsikos G, Pandey NR, Srivastava AK (2013) Insulinomimetic and anti-diabetic effects of zinc. J Inorg Biochem 120:8–17
- Ranasinghe P, Pigera S, Galappatthy P, Katulanda P, Constantine GR (2015) Zinc and diabetes mellitus: understanding molecular mechanisms and clinical implications. DARU J Pharm Sci 23:44
- 78. Tuncay E, Okatan EN, Vassort G, Turan B (2013) ss-blocker timolol prevents arrhythmogenic Ca(2)(+) release and normalizes Ca(2)(+) and Zn(2)(+) dyshomeostasis in hyperglycemic rat heart. PLoS One 8:e71014. https://doi.org/10.1371/journ al.pone.0071014
- Sun Q, Li Q, Zhong W, Zhang J, Sun X, Tan X, Yin X, Sun X, Zhang X, Zhou Z (2014) Dysregulation of hepatic zinc transporters in a mouse model of alcoholic liver disease. Am J Physiol-Gastrointestinal Liver Physiol 307:G313–G322
- Clegg MS, Hanna LA, Niles BJ, Momma TY, Keen CL (2005)
 Zinc deficiency-induced cell death. IUBMB Life 57:661–669
- 81. Maret W (2008) Metallothionein redox biology in the cytoprotective and cytotoxic functions of zinc. Exp Gerontol 43:363–369
- Turan B, Desilets M, Acan LN, Hotomaroglu O, Vannier C, Vassort G (1996) Oxidative effects of selenite on rat ventricular contractility and Ca movements. Cardiovasc Res 32:351–361. https://doi.org/10.1016/0008-6363(96)00071-5
- Klein C, Sunahara RK, Hudson TY, Heyduk T, Howlett AC (2002) Zinc inhibition of cAMP signaling. J Biol Chem 277:11859–11865
- 84. Baltas LG, Karczewski P, Bartel S, Krause EG (1997) The endogenous cardiac sarcoplasmic reticulum Ca2+/calmodulindependent kinase is activated in response to beta-adrenergic stimulation and becomes Ca2+-independent in intact beating hearts. FEBS Lett 409:131-136
- Csermely P, Szamel M, Resch K, Somogyi J (1988) Zinc can increase the activity of protein kinase C and contributes to its binding to plasma membranes in T lymphocytes. J Biol Chem 263:6487–6490
- Tatsumi T, Fliss H (1994) Hypochlorous acid and chloramines increase endothelial permeability: possible involvement of cellular zinc. Am J Phys Heart Circ Phys 267:H1597–H1607
- 87. Oteiza PI (2012) Zinc and the modulation of redox homeostasis. Free Radic Biol Med 53:1748–1759
- Efeovbokhan N, Bhattacharya SK, Ahokas RA, Sun Y, Guntaka RV, Gerling IC, Weber KT (2014) Zinc and the prooxidant heart failure phenotype. J Cardiovasc Pharmacol 64:393



- Krężel A, Maret W (2016) The biological inorganic chemistry of zinc ions. Arch Biochem Biophys 611:3–19
- Hoang BX, Han B, Shaw DG, Nimni M (2016) Zinc as a possible preventive and therapeutic agent in pancreatic, prostate, and breast cancer. Eur J Cancer Prev 25:457

 –461
- Aamodt RL, Rumble WF, Johnston GS, Foster D, Henkin RI (1979) Zinc metabolism in humans after oral and intravenous administration of Zn-69m. Am J Clin Nutr 32:559–569. https:// doi.org/10.1093/ajcn/32.3.559
- 92. Kerchner GA, Canzoniero LM, Yu SP, Ling C, Choi DW (2000) Zn2+ current is mediated by voltage-gated Ca2+ channels and enhanced by extracellular acidity in mouse cortical neurones. J Physiol 528(Pt 1):39–52. https://doi.org/10.111 1/j.1469-7793.2000.00039.x
- Alvarez-Collazo J, Díaz-García CM, López-Medina AI, Vassort G, Alvarez JL (2012) Zinc modulation of basal and β-adrenergically stimulated L-type Ca2+ current in rat ventricular cardiomyocytes: consequences in cardiac diseases. Pflugers Arch 464:459–470. https://doi.org/10.1007/s00424-012-1162-3
- Coudray C, Charlon V, de Leiris J, Favier A (1993) Effect of zinc deficiency on lipid peroxidation status and infarct size in rat hearts. Int J Cardiol 41:109–113. https://doi.org/10.1016/0167-5273(93)90149-b
- 95. Murakami M, Hirano T (2008) Intracellular zinc homeostasis and zinc signaling. Cancer Sci 99:1515–1522. https://doi.org/10.1111/j.1349-7006.2008.00854.x
- Fukunaka A, Fujitani Y (2018) Role of zinc homeostasis in the pathogenesis of diabetes and obesity. Int J Mol Sci 19. https:// doi.org/10.3390/ijms19020476
- Hashemipour M, Kelishadi R, Shapouri J, Sarrafzadegan N, Amini M, Tavakoli N, Movahedian-Attar A, Mirmoghtadaee P, Poursafa P (2009) Effect of zinc supplementation on insulin resistance and components of the metabolic syndrome in prepubertal obese children. Hormones (Athens) 8:279–285. https://doi. org/10.14310/horm.2002.1244
- Rathnayake KM, Silva K, Jayawardena R (2016) Effects of zinc supplementation on obesity: study protocol for a randomized controlled clinical trial. Trials 17:534. https://doi.org/10.1186/ s13063-016-1651-3
- Khorsandi H, Nikpayam O, Yousefi R, Parandoosh M, Hosseinzadeh N, Saidpour A, Ghorbani A (2019) Zinc supplementation improves body weight management, inflammatory biomarkers and insulin resistance in individuals with obesity: a randomized, placebo-controlled, double-blind trial. Diabetol Metab Syndr 11:101. https://doi.org/10.1186/s13098-019-0497-8
- Rios-Lugo MJ, Madrigal-Arellano C, Gaytán-Hernández D, Hernández-Mendoza H, Romero-Guzmán ET (2020) Association of Serum Zinc Levels in overweight and obesity. Biol Trace Elem Res. https://doi.org/10.1007/s12011-020-02060-8
- 101. Hara T, Takeda TA, Takagishi T, Fukue K, Kambe T, Fukada T (2017) Physiological roles of zinc transporters: molecular and genetic importance in zinc homeostasis. J Physiol Sci 67:283–301. https://doi.org/10.1007/s12576-017-0521-4
- 102. Olgar Y, Turan B (2019) A sodium-glucose cotransporter 2 (SGLT2) inhibitor dapagliflozin comparison with insulin shows important effects on Zn(2+)-transporters in cardiomyocytes from insulin-resistant metabolic syndrome rats through inhibition of oxidative stress (1). Can J Physiol Pharmacol 97:528–535. https://doi.org/10.1139/cjpp-2018-0466
- Gong J, Xu J, Bezanilla M, van Huizen R, Derin R, Li M (1999) Differential stimulation of PKC phosphorylation of potassium channels by ZIP1 and ZIP2. Science 285:1565–1569. https://doi. org/10.1126/science.285.5433.1565
- Gaither LA, Eide DJ (2000) Functional expression of the human hZIP2 zinc transporter. J Biol Chem 275:5560–5564. https://doi. org/10.1074/jbc.275.8.5560

- Gaither LA, Eide DJ (2001) The human ZIP1 transporter mediates zinc uptake in human K562 erythroleukemia cells. J Biol Chem 276:22258–22264. https://doi.org/10.1074/jbc.M101772200
- 106. Eide DJ (2006) Zinc transporters and the cellular trafficking of zinc. Biochim Biophys Acta 1763:711–722. https://doi. org/10.1016/j.bbamcr.2006.03.005
- Desouki MM, Geradts J, Milon B, Franklin RB, Costello LC (2007) hZip2 and hZip3 zinc transporters are down regulated in human prostate adenocarcinomatous glands. Mol Cancer 6:37. https://doi.org/10.1186/1476-4598-6-37
- Huang T, Yan G, Guan M (2020) Zinc homeostasis in bone: zinc transporters and bone diseases. Int J Mol Sci 21:1236
- Dufner-Beattie J, Langmade SJ, Wang F, Eide D, Andrews GK (2003) Structure, function, and regulation of a subfamily of mouse zinc transporter genes. J Biol Chem 278:50142–50150. https://doi.org/10.1074/jbc.M304163200
- 110. Liuzzi JP, Cousins RJ (2004) Mammalian zinc transporters. Annu Rev Nutr 24:151–172. https://doi.org/10.1146/annurev.nutr.24.012003.132402
- 111. Huang L, Kirschke CP, Zhang Y, Yu YY (2005) The ZIP7 gene (Slc39a7) encodes a zinc transporter involved in zinc homeostasis of the Golgi apparatus. J Biol Chem 280:15456–15463. https://doi.org/10.1074/jbc.M412188200
- Kirschke CP, Huang L (2003) ZnT7, a novel mammalian zinc transporter, accumulates zinc in the Golgi apparatus. J Biol Chem 278:4096–4102. https://doi.org/10.1074/jbc.M2076 44200
- 113. Taylor KM, Vichova P, Jordan N, Hiscox S, Hendley R, Nicholson RI (2008) ZIP7-mediated intracellular zinc transport contributes to aberrant growth factor signaling in antihormone-resistant breast cancer cells. Endocrinology 149:4912–4920
- 114. Aydemir TB, Chang SM, Guthrie GJ, Maki AB, Ryu MS, Karabiyik A, Cousins RJ (2012) Zinc transporter ZIP14 functions in hepatic zinc, iron and glucose homeostasis during the innate immune response (endotoxemia). PLoS One 7:e48679. https://doi.org/10.1371/journal.pone.0048679
- 115. Sun Q, Zhong W, Zhang W, Li Q, Sun X, Tan X, Sun X, Dong D, Zhou Z (2015) Zinc deficiency mediates alcohol-induced apoptotic cell death in the liver of rats through activating ER and mitochondrial cell death pathways. Am J Physiol Gastrointestinal Liver Physiol 308:G757–G766
- 116. Liu Y, Batchuluun B, Ho L, Zhu D, Prentice KJ, Bhattacharjee A, Zhang M, Pourasgari F, Hardy AB, Taylor KM, Gaisano H, Dai FF, Wheeler MB (2015) Characterization of zinc influx transporters (ZIPs) in pancreatic β cells: roles in regulating cytosolic zinc homeostasis and insulin secretion. J Biol Chem 290:18757–18769. https://doi.org/10.1074/jbc.M115.640524
- 117. Begum NA, Kobayashi M, Moriwaki Y, Matsumoto M, Toyoshima K, Seya T (2002) Mycobacterium bovis BCG cell wall and lipopolysaccharide induce a novel gene, BIGM103, encoding a 7-TM protein: identification of a new protein family having Zn-transporter and Zn-metalloprotease signatures. Genomics 80:630–645. https://doi.org/10.1006/geno.2002.7000
- Croxford TP, McCormick NH, Kelleher SL (2011) Moderate zinc deficiency reduces testicular Zip6 and Zip10 abundance and impairs spermatogenesis in mice. J Nutr 141:359–365
- 119. Ryu M-S, Lichten LA, Liuzzi JP, Cousins RJ (2008) Zinc transporters ZnT1 (Slc30a1), Zip8 (Slc39a8), and Zip10 (Slc39a10) in mouse red blood cells are differentially regulated during erythroid development and by dietary zinc deficiency. J Nutr 138:2076–2083
- Küry S, Dréno B, Bézieau S, Giraudet S, Kharfi M, Kamoun R, Moisan JP (2002) Identification of SLC39A4, a gene involved in acrodermatitis enteropathica. Nat Genet 31:239–240. https:// doi.org/10.1038/ng913



- 121. Li M, Zhang Y, Liu Z, Bharadwaj U, Wang H, Wang X, Zhang S, Liuzzi JP, Chang S-M, Cousins RJ (2007) Aberrant expression of zinc transporter ZIP4 (SLC39A4) significantly contributes to human pancreatic cancer pathogenesis and progression. Proc Natl Acad Sci 104:18636–18641
- 122. Weaver BP, Zhang Y, Hiscox S, Guo GL, Apte U, Taylor KM, Sheline CT, Wang L, Andrews GK (2010) Zip4 (Slc39a4) expression is activated in hepatocellular carcinomas and functions to repress apoptosis, enhance cell cycle and increase migration. PLoS One 5:e13158
- 123. Wang F, Kim B-E, Dufner-Beattie J, Petris MJ, Andrews G, Eide DJ (2004) Acrodermatitis enteropathica mutations affect transport activity, localization and zinc-responsive trafficking of the mouse ZIP4 zinc transporter. Hum Mol Genet 13:563–571
- Chowanadisai W, Lönnerdal B, Kelleher SL (2008) Zip6 (LIV-1) regulates zinc uptake in neuroblastoma cells under resting but not depolarizing conditions. Brain Res 1199:10–19. https://doi.org/10.1016/j.brainres.2008.01.015
- Lopez V, Kelleher SL (2010) Zip6-attenuation promotes epithelial-to-mesenchymal transition in ductal breast tumor (T47D) cells. Exp Cell Res 316:366–375
- 126. Taylor KM, Muraina IA, Brethour D, Schmitt-Ulms G, Nimmanon T, Ziliotto S, Kille P, Hogstrand C (2016) Zinc transporter ZIP10 forms a heteromer with ZIP6 which regulates embryonic development and cell migration. Biochem J 473:2531–2544
- Takatani-Nakase T (2018) Zinc transporters and the progression of breast cancers. Biol Pharm Bull 41:1517–1522
- 128. Colomar-Carando N, Meseguer A, Jutz S, Herrera-Fernández V, Olvera A, Kiefer K, Brander C, Steinberger P, Vicente R (2019) Zip6 transporter is an essential component of the lymphocyte activation machinery. J Immunol 202:441–450
- Shen H, Qin H, Guo J (2009) Concordant correlation of LIV-1 and E-cadherin expression in human breast cancer cell MCF-7. Mol Biol Rep 36:653–659
- 130. Sun P, Wang S, Jiang Y, Tao Y, Tian Y, Zhu K, Wan H, Zhang L, Zhang L (2013) Zip1, Zip2, and Zip8 mRNA expressions were associated with growth hormone level during the growth hormone provocation test in children with short stature. Biol Trace Elem Res 155:11–22
- 131. Matsuura W, Yamazaki T, Yamaguchi-Iwai Y, Masuda S, Nagao M, Andrews GK, Kambe T (2009) SLC39A9 (ZIP9) regulates zinc homeostasis in the secretory pathway: characterization of the ZIP subfamily I protein in vertebrate cells. Biosci Biotechnol Biochem 73:1142–1148
- 132. Thomas P, Dong J (2019) Novel mechanism of endocrine disruption by fungicides through binding to the membrane androgen receptor, ZIP9 (SLC39A9), and antagonizing rapid testosterone induction of the intrinsic apoptotic pathway. Steroids 149:108415. https://doi.org/10.1016/j.steroids.2019.05.007
- 133. Pawan K, Neeraj S, Sandeep K, Ratho RK, Rajendra P (2007) Upregulation of Slc39a10 gene expression in response to thyroid hormones in intestine and kidney. Biochimica et Biophysica Acta (BBA)-Gene Struct Expresn 1769:117–123
- Kagara N, Tanaka N, Noguchi S, Hirano T (2007) Zinc and its transporter ZIP10 are involved in invasive behavior of breast cancer cells. Cancer Sci 98:692

 –697
- 135. Gyulkhandanyan AV, Lu H, Lee SC, Bhattacharjee A, Wijesekara N, Fox JEM, MacDonald PE, Chimienti F, Dai FF, Wheeler MB (2008) Investigation of transport mechanisms and regulation of intracellular Zn2+ in pancreatic α-cells. J Biol Chem 283:10184–10197
- 136. Bin B-H, Bhin J, Takaishi M, Toyoshima K-e, Kawamata S, Ito K, Hara T, Watanabe T, Irié T, Takagishi T (2017) Requirement of zinc transporter ZIP10 for epidermal development: implication of the ZIP10–p63 axis in epithelial homeostasis. Proc Natl Acad Sci 114:12243–12248

- 137. Kelleher SL, Velasquez V, Croxford TP, McCormick NH, Lopez V, MacDavid J (2012) Mapping the zinc-transporting system in mammary cells: molecular analysis reveals a phenotype-dependent zinc-transporting network during lactation. J Cell Physiol 227:1761–1770
- 138. Yu Y, Wu A, Zhang Z, Yan G, Zhang F, Zhang L, Shen X, Hu R, Zhang Y, Zhang K (2013) Characterization of the GufA subfamily member SLC39A11/Zip11 as a zinc transporter. J Nutr Biochem 24:1697–1708
- Chowanadisai W, Graham DM, Keen CL, Rucker RB, Messerli MA (2013) Neurulation and neurite extension require the zinc transporter ZIP12 (slc39a12). Proc Natl Acad Sci U S A 110:9903–9908. https://doi.org/10.1073/pnas.1222142110
- 140. Zhao L, Oliver E, Maratou K, Atanur SS, Dubois OD, Cotroneo E, Chen C-N, Wang L, Arce C, Chabosseau PL (2015) The zinc transporter ZIP12 regulates the pulmonary vascular response to chronic hypoxia. Nature 524:356–360
- 141. Fukada T, Civic N, Furuichi T, Shimoda S, Mishima K, Higashiyama H, Idaira Y, Asada Y, Kitamura H, Yamasaki S (2008) The zinc transporter SLC39A13/ZIP13 is required for connective tissue development; its involvement in BMP/TGF-β signaling pathways. PLoS One 3:e3642
- 142. Fukunaka A, Fukada T, Bhin J, Suzuki L, Tsuzuki T, Takamine Y, Bin B-H, Yoshihara T, Ichinoseki-Sekine N, Naito H (2017) Zinc transporter ZIP13 suppresses beige adipocyte biogenesis and energy expenditure by regulating C/EBP-β expression. PLoS Genet 13:e1006950
- 143. Bin B-H, Hojyo S, Ryong Lee T, Fukada T (2014) Spondy-locheirodysplastic Ehlers-Danlos syndrome (SCD-EDS) and the mutant zinc transporter ZIP13. Rare Dis 2:e974982
- 144. Liuzzi JP, Aydemir F, Nam H, Knutson MD, Cousins RJ (2006) Zip14 (Slc39a14) mediates non-transferrin-bound iron uptake into cells. Proc Natl Acad Sci 103:13612–13617
- Zhao N, Gao J, Enns CA, Knutson MD (2010) ZRT/IRT-like protein 14 (ZIP14) promotes the cellular assimilation of iron from transferrin. J Biol Chem 285:32141–32150
- 146. Hojyo S, Fukada T, Shimoda S, Ohashi W, Bin B-H, Koseki H, Hirano T (2011) The zinc transporter SLC39A14/ZIP14 controls G-protein coupled receptor-mediated signaling required for systemic growth. PLoS One 6:e18059
- Troche C, Beker Aydemir T, Cousins RJ (2016) Zinc transporter Slc39a14 regulates inflammatory signaling associated with hypertrophic adiposity. Am J Physiol Endocrinol Metab 310:E258–E268
- 148. Olgar Y, Durak A, Tuncay E, Bitirim CV, Ozcinar E, Inan MB, Tokcaer-Keskin Z, Akcali KC, Akar AR, Turan B (2018) Increased free Zn(2+) correlates induction of sarco(endo)plasmic reticulum stress via altered expression levels of Zn(2+) -transporters in heart failure. J Cell Mol Med 22:1944–1956. https://doi.org/10.1111/jcmm.13480
- McMahon RJ, Cousins RJ (1998) Mammalian zinc transporters. J Nutr 128:667–670
- Gaither LA, Eide DJ (2001) Eukaryotic zinc transporters and their regulation. Zinc Biochemistry, Physiology, and Homeostasis, Springer, pp 65–84
- 151. Qian J, Xu K, Yoo J, Chen TT, Andrews G, Noebels JL (2011) Knockout of Zn transporters Zip-1 and Zip-3 attenuates seizure-induced CA1 neurodegeneration. J Neurosci 31:97–104
- 152. Inoue K, Matsuda K, Itoh M, Kawaguchi H, Tomoike H, Aoyagi T, Nagai R, Hori M, Nakamura Y, Tanaka T (2002) Osteopenia and male-specific sudden cardiac death in mice lacking a zinc transporter gene, Znt5. Hum Mol Genet 11:1775–1784
- 153. Jin J, Li Z, Liu J, Wu Y, Gao X, He Y (2015) Knockdown of zinc transporter ZIP5 (SLC39A5) expression significantly inhibits human esophageal cancer progression. Oncol Rep 34:1431–1439



- 154. Huang L, Yu YY, Kirschke CP, Gertz ER, Lloyd KK (2007) Znt7 (Slc30a7)-deficient mice display reduced body zinc status and body fat accumulation. J Biol Chem 282:37053–37063
- Huang L, Kirschke CP, Lay Y-AE, Levy LB, Lamirande DE, Zhang PH (2012) Znt7-null mice are more susceptible to dietinduced glucose intolerance and insulin resistance. J Biol Chem 287:33883–33896
- 156. Hogstrand C, Kille P, Nicholson RI, Taylor KM (2009) Zinc transporters and cancer: a potential role for ZIP7 as a hub for tyrosine kinase activation. Trends Mol Med 15:101–111
- Kambe T, Yamaguchi-Iwai Y, Sasaki R, Nagao M (2004) Overview of mammalian zinc transporters. Cell Mol Life Sci CMLS 61:49–68
- 158. Asano N, Kondoh M, Ebihara C, Fujii M, Nakanishi T, Soares MJ, Nakashima E, Tanaka K, Sato M, Watanabe Y (2004) Expression profiles of zinc transporters in rodent placental models. Toxicol Lett 154:45–53
- 159. Olgar Y, Ozdemir S, Turan B (2018) Induction of endoplasmic reticulum stress and changes in expression levels of Zn(2+)transporters in hypertrophic rat heart. Mol Cell Biochem 440:209–219. https://doi.org/10.1007/s11010-017-3168-9
- Cousins RJ, Liuzzi JP, Lichten LA (2006) Mammalian zinc transport, trafficking, and signals. J Biol Chem 281:24085-24089
- 161. Wenzlau JM, Juhl K, Yu L, Moua O, Sarkar SA, Gottlieb P, Rewers M, Eisenbarth GS, Jensen J, Davidson HW (2007) The cation efflux transporter ZnT8 (Slc30A8) is a major autoantigen in human type 1 diabetes. Proc Natl Acad Sci 104:17040–17045
- 162. Tamaki M, Fujitani Y, Hara A, Uchida T, Tamura Y, Takeno K, Kawaguchi M, Watanabe T, Ogihara T, Fukunaka A (2013) The diabetes-susceptible gene SLC30A8/ZnT8 regulates hepatic insulin clearance. J Clin Invest 123:4513–4524
- Pound LD, Sarkar SA, Cauchi S, Wang Y, Oeser JK, Lee CE, Froguel P, Hutton JC, O'Brien RM (2011) Characterization of the human SLC30A8 promoter and intronic enhancer. J Mol Endocrinol 47:251
- 164. Pound LD, Sarkar SA, Ustione A, Dadi PK, Shadoan MK, Lee CE, Walters JA, Shiota M, McGuinness OP, Jacobson DA (2012) The physiological effects of deleting the mouse SLC30A8 gene encoding zinc transporter-8 are influenced by gender and genetic background. PLoS One 7:e40972
- 165. El Muayed M, Billings LK, Raja MR, Zhang X, Park PJ, Newman MV, Kaufman DB, O'Halloran TV, Lowe WL Jr (2010) Acute cytokine-mediated downregulation of the zinc transporter ZnT8 alters pancreatic β-cell function. J Endocrinol 206:159
- 166. Lefebvre B, Vandewalle B, Balavoine A-S, Queniat G, Moerman E, Vantyghem M-C, Le Bacquer O, Gmyr V, Pawlowski V, Kerr-Conte J (2012) Regulation and functional effects of ZNT8 in human pancreatic islets. J Endocrinol 214:225
- Merriman C, Huang Q, Rutter GA, Fu D (2016) Lipid-tuned zinc transport activity of human ZnT8 protein correlates with risk for type-2 diabetes. J Biol Chem 291:26950–26957
- 168. Nicolson TJ, Bellomo EA, Wijesekara N, Loder MK, Baldwin JM, Gyulkhandanyan AV, Koshkin V, Tarasov AI, Carzaniga R, Kronenberger K (2009) Insulin storage and glucose homeostasis in mice null for the granule zinc transporter ZnT8 and studies of the type 2 diabetes—associated variants. Diabetes 58:2070–2083
- 169. Chimienti F, Devergnas S, Favier A, Seve M (2004) Identification and cloning of a β-cell–specific zinc transporter, ZnT-8, localized into insulin secretory granules. Diabetes 53:2330–2337
- 170. Murgia C, Devirgiliis C, Mancini E, Donadel G, Zalewski P, Perozzi G (2009) Diabetes-linked zinc transporter ZnT8 is a homodimeric protein expressed by distinct rodent endocrine cell types in the pancreas and other glands. Nutr Metab Cardiovasc Dis 19:431–439

- Palade GE (1953) An electron microscope study of the mitochondrial structure. J Histochem Cytochem 1:188–211. https:// doi.org/10.1177/1.4.188
- 172. Perez Y, Shorer Z, Liani-Leibson K, Chabosseau P, Kadir R, Volodarsky M, Halperin D, Barber-Zucker S, Shalev H, Schreiber R (2017) SLC30A9 mutation affecting intracellular zinc homeostasis causes a novel cerebro-renal syndrome. Brain 140:928–939
- Sekler I, Sensi SL, Hershfinkel M, Silverman WF (2007) Mechanism and regulation of cellular zinc transport. Molecular medicine, Springer, pp 337–343
- 174. Bosomworth HJ, Thornton JK, Coneyworth LJ, Ford D, Valentine RA (2012) Efflux function, tissue-specific expression and intracellular trafficking of the Zn transporter ZnT10 indicate roles in adult Zn homeostasis. Metallomics 4:771–779
- Brito S, Lee MG, Bin BH, Lee JS (2020) Zinc and its transporters in epigenetics. Mol Cells 43:323–330. https://doi.org/10.14348/ molcells.2020.0026
- 176. Alluri K, Nair KPM, Ghosh S (2020) Differential expression of zinc transporters in functionally contrasting tissues involved in zinc homeostasis. Nucleosides Nucleotides Nucleic Acids 39:615–629. https://doi.org/10.1080/15257770.2019.1670838
- 177. Eide DJ (2020) Transcription factors and transporters in zinc homeostasis: lessons learned from fungi. Crit Rev Biochem Mol Biol 55:88–110. https://doi.org/10.1080/10409238.2020.17420
- 178. Reis BZ, Vieira D, Maynard DDC, Silva DGD, Mendes-Netto RS, Cozzolino SMF (2020) Zinc nutritional status influences ZnT1 and ZIP4 gene expression in children with a high risk of zinc deficiency. J Trace Elem Med Biol 61:126537. https://doi.org/10.1016/j.jtemb.2020.126537
- 179. Turan B (2019) A brief overview from the physiological and detrimental roles of zinc homeostasis via zinc transporters in the heart. Biol Trace Elem Res 188:160–176. https://doi.org/10.1007/s12011-018-1464-1
- Huang T, Yan G, Guan M (2020) Zinc homeostasis in bone: zinc transporters and bone diseases. Int J Mol Sci 21. https://doi. org/10.3390/ijms21041236
- Ohashi W, Fukada T (2019) Contribution of zinc and zinc transporters in the pathogenesis of inflammatory bowel diseases. J Immunol Res 2019:8396878. https://doi.org/10.1155/2019/83968 78
- 182. Martha-Paz AM, Eide D, Mendoza-Cózatl D, Castro-Guerrero NA, Aréchiga-Carvajal ET (2019) Zinc uptake in the Basidiomycota: characterization of zinc transporters in Ustilago maydis. Mol Membr Biol 35:39–50. https://doi.org/10.1080/09687 688.2019.1667034
- Takatani-Nakase T (2018) Zinc transporters and the progression of breast cancers. Biol Pharm Bull 41:1517–1522. https://doi. org/10.1248/bpb.b18-00086
- 184. Ollig J, Kloubert V, Taylor KM, Rink L (2019) B cell activation and proliferation increase intracellular zinc levels. J Nutr Biochem 64:72–79. https://doi.org/10.1016/j.jnutbio.2018.10.008
- 185. Bin BH, Hojyo S, Seo J, Hara T, Takagishi T, Mishima K, Fukada T (2018) The role of the Slc39a family of zinc transporters in zinc homeostasis in skin. Nutrients 10. https://doi.org/10.3390/nu10020219
- 186. Norouzi S, Adulcikas J, Sohal SS, Myers S (2017) Zinc transporters and insulin resistance: therapeutic implications for type 2 diabetes and metabolic disease. J Biomed Sci 24:87. https://doi.org/10.1186/s12929-017-0394-0
- 187. Kessels JE, Wessels I, Haase H, Rink L, Uciechowski P (2016) Influence of DNA-methylation on zinc homeostasis in myeloid cells: regulation of zinc transporters and zinc binding proteins. J Trace Elem Med Biol 37:125–133. https://doi.org/10.1016/j. jtemb.2016.02.003



- 188. Ohashi W, Kimura S, Iwanaga T, Furusawa Y, Irié T, Izumi H, Watanabe T, Hijikata A, Hara T, Ohara O, Koseki H, Sato T, Robine S, Mori H, Hattori Y, Watarai H, Mishima K, Ohno H, Hase K, Fukada T (2016) Zinc transporter SLC39A7/ZIP7 promotes intestinal epithelial self-renewal by resolving ER stress. PLoS Genet 12:e1006349. https://doi.org/10.1371/journal.pgen.1006349
- 189. Fukunaka A, Fukada T, Bhin J, Suzuki L, Tsuzuki T, Takamine Y, Bin BH, Yoshihara T, Ichinoseki-Sekine N, Naito H, Miyatsuka T, Takamiya S, Sasaki T, Inagaki T, Kitamura T, Kajimura S, Watada H, Fujitani Y (2017) Zinc transporter ZIP13 suppresses beige adipocyte biogenesis and energy expenditure by regulating C/EBP-β expression. PLoS Genet 13:e1006950. https://doi.org/10.1371/journal.pgen.1006950
- Takagishi T, Hara T, Fukada T (2017) Recent advances in the role of SLC39A/ZIP zinc transporters in vivo. Int J Mol Sci 18. https://doi.org/10.3390/ijms18122708
- 191. Hara H, Sueyoshi S, Taniguchi M, Kamiya T, Adachi T (2017) Differences in intracellular mobile zinc levels affect susceptibility to plasma-activated medium-induced cytotoxicity. Free Radic Res 51:306–315. https://doi.org/10.1080/10715762.2017.13095
- 192. Eide DJ (2004) The SLC39 family of metal ion transporters. Pflugers Arch 447:796–800. https://doi.org/10.1007/s0042 4-003-1074-3
- Lichten LA, Cousins RJ (2009) Mammalian zinc transporters: nutritional and physiologic regulation. Annu Rev Nutr 29:153– 176. https://doi.org/10.1146/annurev-nutr-033009-083312
- Jeong J, Eide DJ (2013) The SLC39 family of zinc transporters. Mol Asp Med 34:612–619. https://doi.org/10.1016/j.mam.2012.05.011
- Bafaro E, Liu Y, Xu Y, Dempski RE (2017) The emerging role of zinc transporters in cellular homeostasis and cancer. Signal Transduct Target Ther 2:17029. https://doi.org/10.1038/sigtr ans.2017.29
- 196. Myers SA, Nield A, Myers M (2012) Zinc transporters, mechanisms of action and therapeutic utility: implications for type 2 diabetes mellitus. J Nutr Metab 2012:173712. https://doi.org/10.1155/2012/173712
- 197. Kambe T, Tsuji T, Hashimoto A, Itsumura N (2015) The physiological, biochemical, and molecular roles of zinc transporters in zinc homeostasis and metabolism. Physiol Rev 95:749–784. https://doi.org/10.1152/physrev.00035.2014
- Ohashi W, Hara T, Takagishi T, Hase K, Fukada T (2019) Maintenance of intestinal epithelial homeostasis by zinc transporters. Dig Dis Sci 64:2404–2415. https://doi.org/10.1007/s10620-019-05561-2
- Ellis CD, Wang F, MacDiarmid CW, Clark S, Lyons T, Eide DJ (2004) Zinc and the Msc2 zinc transporter protein are required for endoplasmic reticulum function. J Cell Biol 166:325–335. https://doi.org/10.1083/jcb.200401157
- Hogstrand C, Kille P, Nicholson RI, Taylor KM (2009) Zinc transporters and cancer: a potential role for ZIP7 as a hub for tyrosine kinase activation. Trends Mol Med 15:101–111. https://doi.org/10.1016/j.molmed.2009.01.004
- Taylor KM, Vichova P, Jordan N, Hiscox S, Hendley R, Nicholson RI (2008) ZIP7-mediated intracellular zinc transport contributes to aberrant growth factor signaling in antihormone-resistant breast cancer cells. Endocrinology 149:4912–4920. https://doi.org/10.1210/en.2008-0351
- 202. Olgar Y, Degirmenci S, Durak A, Billur D, Can B, Kayki-Mutlu G, Arioglu-Inan E, Turan B (2018) Aging related functional and structural changes in the heart and aorta: MitoTEMPO improves aged-cardiovascular performance. Exp Gerontol 110:172–181. https://doi.org/10.1016/j.exger.2018.06.012

- Grattan BJ, Freake HC (2012) Zinc and cancer: implications for LIV-1 in breast cancer. Nutrients 4:648–675. https://doi. org/10.3390/nu4070648
- Kim JH, Jeon J, Shin M, Won Y, Lee M, Kwak JS, Lee G, Rhee J, Ryu JH, Chun CH, Chun JS (2014) Regulation of the catabolic cascade in osteoarthritis by the zinc-ZIP8-MTF1 axis. Cell 156:730–743. https://doi.org/10.1016/j.cell.2014.01.007
- McAllister BB, Dyck RH (2017) Zinc transporter 3 (ZnT3) and vesicular zinc in central nervous system function. Neurosci Biobehav Rev 80:329–350. https://doi.org/10.1016/j.neubi orev.2017.06.006
- Adlard PA, Parncutt JM, Finkelstein DI, Bush AI (2010) Cognitive loss in zinc transporter-3 knock-out mice: a phenocopy for the synaptic and memory deficits of Alzheimer's disease? J Neurosci 30:1631–1636. https://doi.org/10.1523/jneurosci.5255-09.2010
- 207. Whitfield DR, Vallortigara J, Alghamdi A, Howlett D, Horto-bágyi T, Johnson M, Attems J, Newhouse S, Ballard C, Thomas AJ, O'Brien JT, Aarsland D, Francis PT (2014) Assessment of ZnT3 and PSD95 protein levels in Lewy body dementias and Alzheimer's disease: association with cognitive impairment. Neurobiol Aging 35:2836–2844. https://doi.org/10.1016/j.neurobiolaging.2014.06.015
- 208. Rafalo-Ulinska A, Piotrowska J, Kryczyk A, Opoka W, Sowa-Kucma M, Misztak P, Rajkowska G, Stockmeier CA, Datka W, Nowak G, Szewczyk B (2016) Zinc transporters protein level in postmortem brain of depressed subjects and suicide victims. J Psychiatr Res 83:220–229. https://doi.org/10.1016/j.jpsychires .2016.09.008
- 209. Rafalo A, Zadrozna M, Nowak B, Kotarska K, Wiatrowska K, Pochwat B, Sowa-Kucma M, Misztak P, Nowak G, Szewczyk B (2017) The level of the zinc homeostasis regulating proteins in the brain of rats subjected to olfactory bulbectomy model of depression. Prog Neuro-Psychopharmacol Biol Psychiatry 72:36–48. https://doi.org/10.1016/j.pnpbp.2016.08.009
- Rutter GA (2010) Think zinc: new roles for zinc in the control of insulin secretion. Islets 2:49–50. https://doi.org/10.4161/isl.2.1.10259
- Kawasaki E (2012) ZnT8 and type 1 diabetes. Endocr J 59:531– 537. https://doi.org/10.1507/endocrj.ej12-0069
- Davidson HW, Wenzlau JM, O'Brien RM (2014) Zinc transporter 8 (ZnT8) and β cell function. Trends Endocrinol Metab 25:415–424. https://doi.org/10.1016/j.tem.2014.03.008
- 213. Rutter GA, Chabosseau P, Bellomo EA, Maret W, Mitchell RK, Hodson DJ, Solomou A, Hu M (2016) Intracellular zinc in insulin secretion and action: a determinant of diabetes risk? Proc Nutr Soc 75:61–72. https://doi.org/10.1017/s0029665115003237
- Parsons DS, Hogstrand C, Maret W (2018) The C-terminal cytosolic domain of the human zinc transporter ZnT8 and its diabetes risk variant. FEBS J 285:1237–1250. https://doi.org/10.1111/ febs.14402
- 215. Dwivedi OP, Lehtovirta M, Hastoy B, Chandra V, Krentz NAJ, Kleiner S, Jain D, Richard AM, Abaitua F, Beer NL, Grotz A, Prasad RB, Hansson O, Ahlqvist E, Krus U, Artner I, Suoranta A, Gomez D, Baras A, Champon B, Payne AJ, Moralli D, Thomsen SK, Kramer P, Spiliotis I, Ramracheya R, Chabosseau P, Theodoulou A, Cheung R, van de Bunt M, Flannick J, Trombetta M, Bonora E, Wolheim CB, Sarelin L, Bonadonna RC, Rorsman P, Davies B, Brosnan J, McCarthy MI, Otonkoski T, Lagerstedt JO, Rutter GA, Gromada J, Gloyn AL, Tuomi T, Groop L (2019) Loss of ZnT8 function protects against diabetes by enhanced insulin secretion. Nat Genet 51:1596–1606. https://doi.org/10.1038/s41588-019-0513-9
- Hardy AB, Prentice KJ, Froese S, Liu Y, Andrews GK, Wheeler MB (2015) Zip4 mediated zinc influx stimulates insulin secretion



- in pancreatic beta cells. PLoS One 10:e0119136. https://doi.org/10.1371/journal.pone.0119136
- Norouzi S, Adulcikas J, Sohal SS, Myers S (2018) Zinc stimulates glucose oxidation and glycemic control by modulating the insulin signaling pathway in human and mouse skeletal muscle cell lines. PLoS One 13:e0191727. https://doi.org/10.1371/journal.pone.0191727
- Gray MW, Burger G, Lang BF (1999) Mitochondrial evolution. Science 283:1476–1481. https://doi.org/10.1126/science.283.5407.1476
- Becker T, Vögtle FN, Stojanovski D, Meisinger C (2008) Sorting and assembly of mitochondrial outer membrane proteins. Biochim Biophys Acta 1777:557–563. https://doi.org/10.1016/j.bbabio.2008.03.017
- Zong WX, Rabinowitz JD, White E (2016) Mitochondria and Cancer. Mol Cell 61:667–676. https://doi.org/10.1016/j.molce 1.2016.02.011
- Bruggisser J, Käser S, Mani J, Schneider A (2017) Biogenesis of a mitochondrial outer membrane protein in Trypanosoma brucei: targeting signal and dependence on a unique biogenesis factor. J Biol Chem 292:3400–3410. https://doi.org/10.1074/jbc. M116.755983
- 222. Csordás G, Renken C, Várnai P, Walter L, Weaver D, Buttle KF, Balla T, Mannella CA, Hajnóczky G (2006) Structural and functional features and significance of the physical linkage between ER and mitochondria. J Cell Biol 174:915–921. https://doi.org/10.1083/jcb.200604016
- Rieusset J (2015) Contribution of mitochondria and endoplasmic reticulum dysfunction in insulin resistance: distinct or interrelated roles? Diabetes Metab 41:358–368. https://doi.org/10.1016/j.diabet.2015.02.006
- 224. Sebastián D, Hernández-Alvarez MI, Segalés J, Sorianello E, Muñoz JP, Sala D, Waget A, Liesa M, Paz JC, Gopalacharyulu P, Orešič M, Pich S, Burcelin R, Palacín M, Zorzano A (2012) Mitofusin 2 (Mfn2) links mitochondrial and endoplasmic reticulum function with insulin signaling and is essential for normal glucose homeostasis. Proc Natl Acad Sci U S A 109:5523–5528. https://doi.org/10.1073/pnas.1108220109
- Fawcett DW, McNutt NS (1969) The ultrastructure of the cat myocardium. I. Ventricular papillary muscle. J Cell Biol 42:1– 45. https://doi.org/10.1083/jcb.42.1.1
- 226. Brierley GP, Knight VA (1967) Ion transport by heart mitochondria. X. the uptake and release of Zn2+ and its relation to the energy-linked accumulation of magnesium. Biochemistry 6:3892–3901. https://doi.org/10.1021/bi00864a035
- Saris NE, Niva K (1994) Is Zn2+ transported by the mitochondrial calcium uniporter? FEBS Lett 356:195–198. https://doi.org/10.1016/0014-5793(94)01256-3
- 228. Jiang D, Sullivan PG, Sensi SL, Steward O, Weiss JH (2001) Zn(2+) induces permeability transition pore opening and release of pro-apoptotic peptides from neuronal mitochondria. J Biol Chem 276:47524–47529. https://doi.org/10.1074/jbc.M1088 34200
- 229. Sensi SL, Ton-That D, Weiss JH (2002) Mitochondrial sequestration and Ca(2+)-dependent release of cytosolic Zn(2+) loads in cortical neurons. Neurobiol Dis 10:100–108. https://doi.org/10.1006/nbdi.2002.0493
- Sensi SL, Ton-That D, Sullivan PG, Jonas EA, Gee KR, Kaczmarek LK, Weiss JH (2003) Modulation of mitochondrial function by endogenous Zn2+ pools. Proc Natl Acad Sci U S A 100:6157–6162. https://doi.org/10.1073/pnas.1031598100
- Sensi SL, Ton-That D, Weiss JH, Rothe A, Gee KR (2003) A new mitochondrial fluorescent zinc sensor. Cell Calcium 34:281–284. https://doi.org/10.1016/s0143-4160(03)00122-2
- Burri L, Vascotto K, Fredersdorf S, Tiedt R, Hall MN, Lithgow T (2004) Zim17, a novel zinc finger protein essential for protein

- import into mitochondria. J Biol Chem 279:50243–50249. https://doi.org/10.1074/jbc.M409194200
- 233. Brown AM, Kristal BS, Effron MS, Shestopalov AI, Ullucci PA, Sheu KF, Blass JP, Cooper AJ (2000) Zn2+ inhibits alphaketoglutarate-stimulated mitochondrial respiration and the isolated alpha-ketoglutarate dehydrogenase complex. J Biol Chem 275:13441–13447. https://doi.org/10.1074/jbc.275.18.13441
- Sensi SL, Yin HZ, Carriedo SG, Rao SS, Weiss JH (1999) Preferential Zn2+ influx through Ca2+-permeable AMPA/kainate channels triggers prolonged mitochondrial superoxide production. Proc Natl Acad Sci U S A 96:2414–2419. https://doi.org/10.1073/pnas.96.5.2414
- 235. Zhang Y, Xing F, Zheng H, Xi J, Cui X, Xu Z (2013) Roles of mitochondrial Src tyrosine kinase and zinc in nitric oxide-induced cardioprotection against ischemia/reperfusion injury. Free Radic Res 47:517–525. https://doi.org/10.3109/10715762.2013.796044
- 236. Sharpley MS, Hirst J (2006) The inhibition of mitochondrial complex I (NADH:ubiquinone oxidoreductase) by Zn2+. J Biol Chem 281:34803–34809. https://doi.org/10.1074/jbc.M607389200
- Dineley KE, Richards LL, Votyakova TV, Reynolds IJ (2005)
 Zinc causes loss of membrane potential and elevates reactive oxygen species in rat brain mitochondria. Mitochondrion 5:55–65. https://doi.org/10.1016/j.mito.2004.11.001
- Sharaf MS, Stevens D, Kamunde C (2017) Zinc and calcium alter the relationship between mitochondrial respiration, ROS and membrane potential in rainbow trout (Oncorhynchus mykiss) liver mitochondria. Aquat Toxicol 189:170–183. https://doi. org/10.1016/j.aquatox.2017.06.005
- Rasola A, Bernardi P (2011) Mitochondrial permeability transition in Ca(2+)-dependent apoptosis and necrosis. Cell Calcium 50:222–233. https://doi.org/10.1016/j.ceca.2011.04.007
- Kelleher SL, Lönnerdal B (2003) Zn transporter levels and localization change throughout lactation in rat mammary gland and are regulated by Zn in mammary cells. J Nutr 133:3378–3385. https://doi.org/10.1093/jn/133.11.3378
- Turan B (2003) Zinc-induced changes in ionic currents of cardiomyocytes. Biol Trace Elem Res 94:49–60. https://doi. org/10.1385/bter:94:1:49
- 242. Traynelis SF, Burgess MF, Zheng F, Lyuboslavsky P, Powers JL (1998) Control of voltage-independent zinc inhibition of NMDA receptors by the NR1 subunit. J Neurosci 18:6163–6175. https://doi.org/10.1523/jneurosci.18-16-06163.1998
- Gilly WF, Armstrong CM (1982) Slowing of sodium channel opening kinetics in squid axon by extracellular zinc. J Gen Physiol 79:935–964. https://doi.org/10.1085/jgp.79.6.935
- 244. Gao H, Boillat A, Huang D, Liang C, Peers C, Gamper N (2017) Intracellular zinc activates KCNQ channels by reducing their dependence on phosphatidylinositol 4,5-bisphosphate. Proc Natl Acad Sci U S A 114:E6410–e6419. https://doi.org/10.1073/ pnas.1620598114
- 245. Medvedeva YV, Weiss JH (2014) Intramitochondrial Zn2+ accumulation via the Ca2+ uniporter contributes to acute ischemic neurodegeneration. Neurobiol Dis 68:137–144. https://doi.org/10.1016/j.nbd.2014.04.011
- 246. Holmuhamedov EL, Wang L, Terzic A (1999) ATP-sensitive K+ channel openers prevent Ca2+ overload in rat cardiac mitochondria. J Physiol 519(Pt 2):347–360. https://doi.org/10.1111/j.1469-7793.1999.0347m.x
- Gunter TE, Buntinas L, Sparagna G, Eliseev R, Gunter K (2000) Mitochondrial calcium transport: mechanisms and functions. Cell Calcium 28:285–296. https://doi.org/10.1054/ceca.2000.0168
- Balaban RS (2002) Cardiac energy metabolism homeostasis: role of cytosolic calcium. J Mol Cell Cardiol 34:1259–1271. https:// doi.org/10.1006/jmcc.2002.2082



- 249. Argaud L, Gateau-Roesch O, Raisky O, Loufouat J, Robert D, Ovize M (2005) Postconditioning inhibits mitochondrial permeability transition. Circulation 111:194–197. https://doi.org/10.1161/01.Cir.0000151290.04952.3b
- 250. Salazar G, Huang J, Feresin RG, Zhao Y, Griendling KK (2017) Zinc regulates Nox1 expression through a NF-κB and mitochondrial ROS dependent mechanism to induce senescence of vascular smooth muscle cells. Free Radic Biol Med 108:225–235. https://doi.org/10.1016/j.freeradbiomed.2017.03.032
- 251. Hojyo S, Fukada T (2016) Zinc transporters and signaling in physiology and pathogenesis. Arch Biochem Biophys 611:43–50. https://doi.org/10.1016/j.abb.2016.06.020
- Chabosseau P, Rutter GA (2016) Zinc and diabetes. Arch Biochem Biophys 611:79–85. https://doi.org/10.1016/j. abb.2016.05.022
- Maret W (2017) Zinc in cellular regulation: the nature and significance of "zinc signals". Int J Mol Sci 18. https://doi. org/10.3390/ijms18112285
- Fukada T, Kambe T (2018) Welcome to the world of zinc signaling. Int J Mol Sci 19. https://doi.org/10.3390/ijms19030785
- Lin W, Li D, Cheng L, Li L, Liu F, Hand NJ, Epstein JA, Rader DJ (2018) Zinc transporter Slc39a8 is essential for cardiac ventricular compaction. J Clin Invest 128:826–833. https://doi.org/10.1172/jci96993
- 256. Turan B, Billur D, Olgar Y (2019) Zinc signaling in aging heart function. In: Fukada T, Kambe T (eds) Zinc Signaling. Springer, Singapore, pp 139–164
- Seo YA, Lopez V, Kelleher SL (2011) A histidine-rich motif mediates mitochondrial localization of ZnT2 to modulate mitochondrial function. Am J Physiol Cell Physiol 300:C1479– C1489. https://doi.org/10.1152/ajpcell.00420.2010
- 258. Cho HM, Ryu JR, Jo Y, Seo TW, Choi YN, Kim JH, Chung JM, Cho B, Kang HC, Yu SW, Yoo SJ, Kim H, Sun W (2019) Drp1-Zip1 interaction regulates mitochondrial quality surveillance system. Mol Cell 73:364–376. https://doi.org/10.1016/j.molce l.2018.11.009

- 259. Pivovarova NB, Stanika RI, Kazanina G, Villanueva I, Andrews SB (2014) The interactive roles of zinc and calcium in mitochondrial dysfunction and neurodegeneration. J Neurochem 128:592–602. https://doi.org/10.1111/jnc.12489
- Bonanni L, Chachar M, Jover-Mengual T, Li H, Jones A, Yokota H, Ofengeim D, Flannery RJ, Miyawaki T, Cho CH, Polster BM, Pypaert M, Hardwick JM, Sensi SL, Zukin RS, Jonas EA (2006) Zinc-dependent multi-conductance channel activity in mitochondria isolated from ischemic brain. J Neurosci 26:6851–6862. https://doi.org/10.1523/jneurosci.5444-05.2006
- Medvedeva YV, Lin B, Shuttleworth CW, Weiss JH (2009)
 Intracellular Zn2+ accumulation contributes to synaptic failure, mitochondrial depolarization, and cell death in an acute slice oxygen-glucose deprivation model of ischemia. J Neurosci 29:1105–1114. https://doi.org/10.1523/jneurosci.4604-08.2009
- Malaiyandi LM, Honick AS, Rintoul GL, Wang QJ, Reynolds IJ (2005) Zn2+ inhibits mitochondrial movement in neurons by phosphatidylinositol 3-kinase activation. J Neurosci 25:9507– 9514. https://doi.org/10.1523/jneurosci.0868-05.2005
- 263. Devinney MJ, Malaiyandi LM, Vergun O, DeFranco DB, Hastings TG, Dineley KE (2009) A comparison of Zn2+- and Ca2+-triggered depolarization of liver mitochondria reveals no evidence of Zn2+-induced permeability transition. Cell Calcium 45:447-455. https://doi.org/10.1016/j.ceca.2009.03.002
- 264. Wang F, Kim B-E, Petris MJ, Eide DJ (2004) The mammalian Zip5 protein is a zinc transporter that localizes to the basolateral surface of polarized cells. J Biol Chem 279:51433–51441
- 265. Olgar Y, Turan B (2019) A sodium-glucose cotransporter 2 (SGLT2) inhibitor dapagliflozin comparison with insulin shows important effects on Zn2+-transporters in cardiomyocytes from insulin-resistant metabolic syndrome rats through inhibition of oxidative stress. Can J Physiol Pharmacol 97:528-535

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

